

Cleveland Clinic Laboratories

Technical Update • August 2017

Cleveland Clinic Laboratories is dedicated to keeping you updated and informed about recent testing changes. This Technical Update is provided on a monthly basis to notify you of any changes to the tests in our catalog.

Recently changed tests are bolded, and they could include revisions to methodology, reference range, days performed, or CPT code. Deleted tests and new tests are listed separately. For your convenience, tests are listed alphabetically and order codes are provided.

To compare the new information with previous test information, refer to the online Test Directory at clevelandcliniclabs. com. Test information is updated in the online Test Directory on the Effective Date stated in the Technical Update. Please update your database as necessary.

For additional detail, contact Client Services at 216.444.5755 or 800.628.6816, or via email at clientservices@ccf.org.

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3	17-Hydroxyprogesterone, Urine														
3	50 gram, non-fasting, 1-hour, gestational glucose screen														
3	75 gram, fasting, 2-hour, non-gestational glucose tolerance														
3	75 gram, fasting, 5-hour, non-gestational glucose tolerance														
3	100 gram, fasting, 3-hour gestational glucose tolerance confirmation														
3	Alcohol, Blood Confirmation														
4	Alcohols														
19	Amniotic Bilirubin Scan														
19	Bupivacaine														
4	Candida albicans Abs, IgA, IgG, IgM														
4	CAR Autoantibody														l
5	Cortisol, Saliva														ĺ
15	Cytochrome P450 2D6 (CYP2D6) Geno														
19	Cyto P450 2D6 Geno														
16	ERBB2 (HER2/neu) Gene Amplification by FISH, Gastric Tissue														
19	FISH for BK Virus														
19	Gastric Analysis														
16–17	Homocysteine, Total, Urine														
19	Homocystine, Urine Quantitative														

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	5	Hu Autoantibody													
	5	Hypersensitivity Pneumonitis II													
	6	Isopropanol													
	19	Leucine Aminopeptidase (LAP), Serum													
	6,19	Lipid Associated Sialic Acid													
	6–7	Manganese, Urine													
	7	Methanol													
	8	Multifocal Neuropathy Evaluation													
	8	Neoencephalitis Paraneoplastic Profile with Recombx													
	8	Neopterin													
	9	Neosensory Neuropathy Paraneoplastic Profile													
	9–10	Nickel, Urine 24 Hour													
	10	Pancreatic Elastase, Fecal													
	10	Recombx CV2 Autoantibody Test													
	10	Recombx MaTa Autoantibody Test													
	17	Reticulin Antibodies, Serum													
	19	Reticulin Antibody, IgA with reflex to Titer													
	11	RI Autoantibody													
	11	SensoriMotor Neuropathy Profile Complete													
	12	Sensory Neuropathy Profile xp													
	12	Thallium, Blood													
	13	Thallium, Urine													
	13	Toluene, Blood													
	18	Toxicology Screen w/ confirmation, Urine													
	13	Valproic Acid, Total and Free													
	19	Xylose Absorption Test (Adult)													
	19	Xylose Absorption Test, Child													
	14	Yo Autoantibody													

Test Changes

Test Name	Order Code	Change	Effective Date
17-Hydroxypro- gesterone, Urine	U170HP	Special Information: Patient should not be on any corticosteroid, ACTH, estrogen, or gonadotropin medications, if possible, for at least 48 hours prior to collection of specimen. Separate specimens must be submitted when multiple tests are ordered. No special preservatives required. Submit total volume. Note: Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible. This test is not New York DOH approved at any laboratory. An approved non-permitted laboratory (NPL) form must accompany specimen. Specimen Requirement: 5 mL 24-hour urine (well-mixed) in a clean container; Minimum: 1 mL; Refrigerate during collection; Patient should not be on any corticosteroid, ACTH, estrogen, or gonadotropin medications, if possible, for at least 48 hours prior to start of urine collection; Transfer 5 mL aliquot to standard transport tube; Submit total volume; Transport on dry ice is preferred; Separate specimens must be submitted when multiple tests are ordered; Frozen Stability: Ambient: 1 hour Refrigerated: 4 days Frozen: 6 months Days Performed: Varies Reported: 7–10 days	8/2/17
50 gram, non- fasting, 1-hour, gestational glucose screen	GLTGST	Reference Range: Glucose Screen, Pregnancy: 74–134 mg/dL	9/28/17
75 gram, fasting, 2-hour, non- gestational glucose tolerance	GTNG2	Reference Range: Fasting: 74–99 mg/dL 1 Hour: See comment 2 Hour: 74–139 mg/dL	9/28/17
75 gram, fasting, 5-hour, non- gestational glucose tolerance	GTNG5	Reference Range: Fasting: 74–99 mg/dL 1 Hour: See comment 2 Hour: 74–139 mg/dL 3 Hour: See comment 4 Hour: See comment 5 Hour: See comment	9/28/17
100 gram, fasting, 3-hour gestational glucose tolerance confirmation	GTGST3	Reference Range: Glucose GST, Fasting: 74–94 mg/dL Glucose GST, 1 Hr: 74–179 mg/dL Glucose GST, 2 Hr: 74–154 mg/dL Glucose GST, 3 Hr: 74–139 mg/dL	9/28/17
Alcohol, Blood Confirmation	BALCO	Special Information: For medical purposes only. Timing of specimen collection: Dependent on time of exposure, test upon presentation to hospital. Cap tube tightly to minimize alcohol loss. Do NOT freeze whole blood. Unacceptable conditions: Whole blood, plasma separator tubes (PST), serum separator tubes (SST). This test is New York DOH approved. Specimen Requirement: 2 mL serum from a red top tube with no additive; Minimum: 0.5 mL; Separate serum from cells ASAP or within 2 hours of collection; Transfer 2 mL serum to a standard transport tube; Cap tube tightly to minimize alcohol loss; Do NOT draw serum separator tubes (SST); Refrigerated *OR* 2 mL plasma from a sodium or lithium heparin green top tube; Minimum: 0.5 mL; Separate plasma from cells ASAP or within 2 hours of collection; Transfer 2 mL plasma to a standard transport tube; Cap tube tightly to minimize alcohol loss; Do NOT draw plasma separator tubes (PST); Refrigerated *OR* 2 mL plasma from a potassium oxalate/sodium fluoride (gray) tube; Minimum: 0.5 mL; Separate plasma from cells ASAP or within 2 hours of collection; Transfer 2 mL plasma to a standard transport tube; Cap tube tightly to minimize alcohol loss; Do NOT draw plasma separator tubes (PST); Refrigerated *OR* 2 mL plasma from a potassium oxalate/sodium fluoride (gray) tube; Minimum: 0.5 mL; Separate plasma to a standard transport tube; Cap tube tightly to minimize alcohol loss; Do NOT draw plasma separator tubes (PST); Refrigerated *OR* 2 mL plasma from an EDTA lavender top tube; Minimum: 0.5 mL; Separate plasma from cells ASAP or within 2 hours of collection; Transfer 2 mL plasma to a standard transport tube; Cap tube tightly to minimize alcohol loss; Do NOT draw plasma separator tubes (PST); Refrigerated Days Performed: Sunday–Saturday Reported: 2–4 days	8/21/17

Test Name	Order Code	Change	Effective Date
Alcohols	ALCOS	Special Information: Patient Prep: For medical purposes only. Timing of specimen collection: Dependent on time of exposure, test upon presentation to hospital. Cap tube tightly to minimize alcohol loss. Unacceptable conditions: Whole blood, plasma separator tubes (PST), serum separator tubes (SST). This test is New York DOH approved. Clinical Information: Use to identify ethanol, methanol, isopropanol or acetone ingestion. Stability: Ambient: 1 week: After separation from cells (if tightly capped) Refrigerated: 2 weeks: After separation from cells (if tightly capped) Frozen: 1 month: After separation from cells Days Performed: Sunday–Saturday Reported: 2–4 days	8/21/17
Candida albicans Abs, IgA, IgG, IgM	CNDAGM	Special Information: Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible. Clinical Information: Limited clinical utility. Reference Range: Candida albicans Ab, IgA 0.88 EV or less: Negative-No significant level of detectable Candida albicans antibody 0.89-0.99 EV: Equivocal: Questionable presence of antibodies; Repeat testing in 10-14 days may be helpful 1.00 EV or greater: Positive: Antibody to Candida albicans detected, which may indicate a current or past infection Candida albicans Ab, IgG 0.88 EV or less: Negative-No significant level of detectable Candida albicans antibody 0.89-0.99 EV: Equivocal-Questionable presence of antibodies; Repeat testing in 10-14 days may be helpful 1.00 EV or greater: Positive-Antibody to Candida albicans detected, which may indicate a current or past infection Candida albicans Ab, IgG 0.89-0.99 EV: Equivocal-Questionable presence of antibodies; Repeat testing in 10-14 days may be helpful 1.00 EV or greater: Positive-Antibody to Candida albicans detected, which may indicate a current or past infection Candida albicans Ab, IgM 0.88 EV or less: Negative-No significant level of detectable Candida albicans antibody 0.89-0.99 EV: Equivocal-Questionable presence of antibodies; Repeat testing in 10-14 days may be helpful 1.00 EV or greater: Positive-Antibody to Candida albicans detected, which may indicate a current or past infection 0.89-0.99 EV: Equivocal-Questionable presence of antibodies; Repeat testing in 10-14 days may be helpful 1.00 EV or greater: Positive-Antibody to Candida albicans detected, which may indicate a current or past infection	8/21/17
CAR Autoantibody	CARAB	Special Information: Serum must be separated from cells within 48 hours of collection. Whole blood is not an acceptable specimen. Clinical Information: Anti-recoverin antibodies can be detected in patients with cancer-associated retinopathy (CAR). Specimen Requirement: 2 mL serum from a serum separator (gold) tube; Minimum: 0.5 mL; Serum must be separated from cells within 48 hours of collection; Refrigerated *OR* 2 mL serum from a red top tube with no additive; Minimum: 0.5 mL; Serum must be separated from cells within 48 hours of collection; Refrigerated *OR* 2 mL cerebrospinal fluid (CSF) in a sterile container; Minimum: 0.5 mL; Collect in sterile, leak-proof container; Refrigerated Stability: Ambient: 72 hours Refrigerated: 28 days Frozen: 28 days Methodology: Automated Nanoliter Scale Immunoassay Reference Range: Serum: < 1:50 CSF: < 1:1 Days Performed: Tuesday, Friday Reported: 4–6 days CPT: 83520 x 1	8/28/17

Test Name	Order Code	Change	Effective Date
Cortisol, Saliva	SCORT	 Special Information: Collection instructions: 1. Do not brush teeth before collecting specimen. 2. Do not eat or drink for 15 minutes prior to specimen collection. 3. Collect specimen between 11 p.m. and midnight, and record collection time. 4. To use Salivette (preferred collection unit): a. Remove top cap of container to expose swab. b. Place swab directly into mouth by tipping container so that swab falls into mouth. Do not touch swab with fingers. c. Keep swab in mouth for approximately two minutes. Roll swab in mouth, do not chew swab. d. Place swab back into its container without touching, and replace the cap. e. Record collection time. Refrigerate. If multiple specimens are collected, submit each vial under a separate order. This test is New York State approved. Clinical Information: Screening for Cushing syndrome. Diagnosis of Cushing syndrome in patients presenting with symptoms or signs suggestive of the disease. CAUTION: Acute stress (including hospitalization and surgery), alcoholism, depression, and many drugs (e.g., exogenous glucocorticoids, anticonvulsants) can obliterate normal diurnal variation, affect response to suppression/stimulation tests and cause elevated cortisol levels. Cortisol levels may be increased in pregnancy and with exogenous estrogens. Specimen Requirement: 1.5 mL saliva in a SARSTEDT Salivette (T514) saliva collection kit; Minimum: 0.6 mL; Refer to Special Info for Salivette collection instructions; Do not brush teeth before collecting specimen; Do not eat or drink for 15 minutes prior to collection; Collect specimen between 11 p.m. and midnight (preferred time), and record collection date and time on container; Refrigerated Stability: Ambient: 1 week Refrigerated: 1 week Frozen: 60 days 	Effective immediately
Hu Autoantibody	ANTIHU	Special Information: Serum must be separated from cells within 48 hours of collection. Clinical Information: Anti-Hu (ANNA1) antibody is found in 5–10% of patients with small cell carcinoma of the lung and is associated with paraneoplastic encephalomyelitis and sensory neuropathy. This semi-quantitative assay may be useful in monitoring therapeutic responses in select cases. Specimen Requirement: 2 mL serum from a red top tube with no additive; Minimum: 0.5 mL; Serum must be separated from cells within 48 hours of collection; Refrigerated *OR* 2 mL serum from a serum separator (gold) tube; Minimum: 0.5 mL; Serum must be separated from cells within 48 hours of collection; Refrigerated *OR* 2 mL cerebrospinal fluid (CSF) in a sterile container; Minimum: 0.5 mL; Collect in sterile, leak-proof container; Refrigerated Stability: Ambient: 72 hours Refrigerated: 28 days Frozen: 28 days Methodology: Automated Nanoliter Scale Immunoassay Reference Range: Serum: < 1:100 CSF: < 1:1 Days Performed: Monday–Friday Reported: 4–6 days CPT: 83520 x 1	8/28/17
Hypersensitivity Pneumonitis II	HYPNE2	Specimen Requirement: 3 mL serum from a serum separator (gold) tube; Minimum: 2 mL; Refrigerated *OR* 3 mL serum from a red top tube with no additive; Minimum: 2 mL; Refrigerated Methodology: Double Diffusion (DD) ImmunoCAP Days Performed: Sunday, Wednesday, Friday Reported: 8–9 days	Effective immediately

Test Name	Order Code	Change	Effective Date
Isopropanol	ISOPRO	Special Information: Patient Prep: For medical purposes only. Timing of specimen collection: Dependent on time of exposure, test upon presentation to hospital. Do NOT freeze whole blood. Cap tube tightly to minimize alcohol loss. Unacceptable conditions: Whole blood, plasma separator tubes (PST), serum separator tubes (SST). This test is New York DOH approved. Clinical Information: Monitor exposure to isopropanol. Toxic concentrations may cause nausea, dizziness, central nervous system depression and coma. Specimen Requirement: 3 mL serum from a red top tube with no additive; Minimum: 0.5 mL; Separate serum from cells ASAP or within 2 hours of collection;	8/21/17
		Transfer 3 mL serum to a standard transport tube; Cap tube tightly to minimize alcohol loss; Do not draw serum separator tubes (SST); Refrigerated *OR* 3 mL plasma from a potassium oxalate/sodium fluoride (gray) tube; Minimum: 0.5 mL; Separate plasma from cells ASAP or within 2 hours of	
		collection; Transfer 3 mL plasma to a standard transport tube; Cap tube tightly to minimize alcohol loss; Do not draw plasma separator tubes (PST); Refrigerated Reference Range: Isopropanol No therapeutic range; Assay detection limit: 5 mg/dL Acetone	
		No therapeutic range; Assay detection limit: 5 mg/dL	
		Reported: 2–4 days	
Lipid Associated Sialic Acid	LIPSIA	Special Information: If tube other than a gel-barrier tube is used, transfer separated serum or plasma to a plastic transport tube.	8/21/17
Sialic Acid		Clinical Information: Lipid Associated Sialic Acid is a useful adjunct in the management of a variety of malignancies. It is generally used in conjunction with other tumor markers. Results for this test are for investigational purposes only by the assay's manufacturer. The performance characteristics of this product have not been established. Results should not be used as a diagnostic procedure without confirmation of the diagnosis by another medically established diagnostic product or procedure. Values obtained with different assay methods should not be used interchangeably in serial testing. It is recommended that only one assay method be used consistently to monitor each patient's course of therapy. This procedure does not provide serial monitoring; it is intended for one-time use only.	
		•OR* 2 mL serum from a red top tube with no additive: Minimum: 0.1 mL;	
		Separate serum from cells and transfer to a standard plastic aliquot tube; Ambient *OR* 2 mL plasma from an EDTA lavender top tube; Minimum: 0.1 mL; Separate	
		plasma from cells and transfer to a standard plastic aliquot tube; Ambient Stability: Ambient: 14 days Refrigerated: 14 days Frozen: 14 days; Stable for 3 freeze/thaw cycles	
		Reference Range: < 20 mg/dL	
		Days Performed: Tuesday, Thursday Reported: 2–4 days	
Manganese, Urine	UMANG	Note: There is a clinically significant charting name change associated with this test. Includes: Creatinine mg/dL Creatinine mg/day Manganese Urine_per volume	8/21/17
		Manganese, Urine-per 24h Manganese, Urine-ratio to CRT (continued on page 7)	

Test Name	Order Code	Change	Effective Date
Manganese, Urine (continued from page 6)		Special Information: Collection volume MUST be indicated. Diet, medication and nutritional supplements may introduce interfering substances. Patients should be encouraged to discontinue nutritional supplements, vitamins, minerals and nonessential over-the-counter medications (upon the advice of their physician). High concentrations of iodine may interfere with elemental testing. Abstinence from iodine-containing medications or contrast agents for at least one month prior to collecting specimens is recommended. ARUP studies indicate that refrigeration of urine alone, during and after collection, preserves specimens adequately, if tested within 14 days of collection. Urine collected within 48 hours after administration of a gadolinium (Gd) containing contrast media (may occur with MRI studies) and acid preserved samples are unacceptable. Specimens contaminated with blood or fecal material and specimens transported in a non-trace element free transport tube (with the exception of the original device) are also unacceptable. This test is New York DOH approved.	
		Clinical Information: This assay provides limited utility in determining manganese exposure. Whole blood measurements are recommended for determining recent or active exposure.	
		Specimen Requirement: 8 mL 24-hour urine (well-mixed) in a clean container; Minimum: 1 mL; Refrigerate during collection; Collect in a plastic container; Aliquot into a trace metal free transport tube (ARUP #43116); Record total volume and collection time interval on tube and requisition; Refrigerated	
		OR 8 mL random urine in a clean container; Minimum: 1 mL; Refrigerate during collection; Collect in a plastic container; Aliquot into a trace metal free transport tube (ARUP #43116); Record total volume on tube and requisition; Refrigerated	
		Reference Range: Manganese, Urine-ratio to CRT: 0.0–0.9 µg/g crt Manganese, Urine-per volume: 0.0–0.9 µg/L Manganese, Urine-per 24h: 0.0–2.4 µg/d Creatinine mg/day: Refer to report	
		Days Performed: Sunday–Saturday Reported: 2–6 days	
Methanol	METHOL	Special Information: Patient Prep: For medical purposes only. Timing of specimen collection: Dependent on time of exposure, test upon presentation to hospital. Do NOT freeze whole blood. Cap tube tightly to minimize alcohol loss. Unacceptable conditions: Whole blood, plasma separator tubes (PST) and serum separator tubes (SST). This test is New York DOH approved.	8/21/17
		Clinical Information: Monitor exposure to methanol. Toxic concentrations may cause intoxication, metabolic acidosis, ocular toxicity, central nervous system (CNS) depression and fatality if patients do not receive medical treatment.	
		Specimen Requirement: 3 mL serum from a red top tube with no additive; Minimum: 0.5 mL; Do not use serum separator tubes (SST); Separate serum from cells ASAP or within 2 hours of collection; Transfer 3 mL serum to a standard transport tube; Cap tube tightly to minimize alcohol loss; Refrigerated	
		OR 3 mL plasma from a sodium or lithium heparin green top tube; Minimum: 0.5 mL; Do not use plasma separator tubes (PST); Separate plasma from cells ASAP or within 2 hours of collection; Transfer 3 mL plasma to a standard transport tube; Cap tube tightly to minimize alcohol loss; Refrigerated	
		OR 3 mL plasma from a potassium oxalate/sodium fluoride (gray) tube; Minimum: 0.5 mL; Do not use plasma separator tubes (PST); Separate plasma from cells ASAP or within 2 hours of collection; Transfer 3 mL plasma to a standard transport tube; Cap tube tightly to minimize alcohol loss; Refrigerated	
		OR 3 mL plasma from an EDTA lavender top tube; Minimum: 0.5 mL; Do not use plasma separator tubes (PST); Separate plasma from cells ASAP or within 2 hours of collection; Transfer 3 mL plasma to a standard transport tube; Cap tube tightly to minimize alcohol loss; Refrigerated	
		Days Performed: Sunday-Saturday	
		Reported: ∠–4 days	

Test Name	Order Code	Change	Effective Date
Multifocal Neuropathy Evaluation	MULNEU	Special Information: Informed consent is required. Whole blood collected in a red top or serum separator tube (SST) is not acceptable. Hemolyzed, icteric or lipemic specimens will be rejected. Serum must be separated from cells within 48 hours of collection. Clinical Information: For detection of a mutation in the gene causing HNPP. Detect presence of anti-GM1, anti-GD1a, anti-Asialo GM1, and anti-GD1b antibodies. Specimen Requirement: 8 mL whole blood in an EDTA lavender top tube; Minimum: 6 mL; THIS ASSAY REQUIRES MULTIPLE SPECIMEN TYPES; Collect 2 EDTA lavender top tubes to ensure adequate specimen volume; Send to Cleveland Clinic Laboratories on the day of collection; Refrigerated *AND* 2 mL serum from a serum separator (gold) tube; Minimum: 0.5 mL; Serum must be separated from cells within 48 hours of collection; Must send refrigerated; Refrigerated *OR* 8 mL whole blood in an EDTA lavender top tube; Minimum: 6 mL; THIS ASSAY REQUIRES MULTIPLE SPECIMEN TYPES; Collect 2 EDTA lavender top tubes to ensure adequate specimen volume; Refrigerated *AND* 2 mL serum from a red top tube with no additive; Minimum: 0.5 mL; Serum must be separated from cells within 48 hours of collection; Must send refrigerated; Refrigerated *AND* 2 mL serum from a red top tube with no additive; Minimum: 0.5 mL; Serum must be separated from cells within 48 hours of collection; Must send refrigerated; Refrigerated Stability: Ambient: Whole blood: 10 days; Serum: 3 days Refrigerated: Whole blood: 10 days; Serum: 3 days Frozen: Whole blood: 10 days; Serum: 4 months Methodology: Enzyme-Linked Immunosorbent Assay (ELISA) Multiplex Ligation-dependent Probe Amplification Reference Range: Refer to report Days Performed: Tuesday Reported: 15–22 days CPT: 81324 x 1, 83520 x 4	8/28/17
Neoencephalitis Paraneoplastic Profile with Recombx	CEPHAL	Special Information: Whole blood is not an acceptable specimen. Serum must be separated from cells within 48 hours of collection. Clinical Information: Diseases tested for include Paraneoplastic Neurological Syndrome, Autoimmune Encephalitis, and Stiff Person Syndrome. Specimen Requirement: 2 mL serum from a red top tube with no additive; Minimum: 0.5 mL; Serum must be separated from cells within 48 hours of collection; Refrigerated *OR* 2 mL serum from a serum separator (gold) tube; Minimum: 0.5 mL; Serum must be separated from cells within 48 hours of collection; Refrigerated Stability: Ambient: 72 hours Refrigerated: 28 days Frozen: 28 days Methodology: Automated Nanoliter Scale Immunoassay Enzyme-Linked Immunosorbent Assay (ELISA) Immunofluorescence Radioimmunoassay (RIA) Reference Range: Refer to report Days Performed: Monday, Thursday Reported: 8–11 days CPT: 83519 x 1, 83520 x 4, 86255 x 3, 86341 x 1	8/28/17
Neopterin	NEOPT	Reference Range: Adults: < 2.5 ng/mL	9/27/17

Test Name	Order Code	Change	Effective Date
Neosensory Neuropathy Paraneoplastic Profile	NEOSEN	Special Information: Serum must be separated from cells within 48 hours of collection. Whole blood is not an acceptable specimen.	8/28/17
		Clinical Information: Diseases tested for include Paraneoplastic Neurological Syndrome and Stiff Person Syndrome.	
		Specimen Requirement: 2 mL serum from a red top tube with no additive; Minimum: 0.5 mL; Serum must be separated from cells within 48 hours of collection; Refrigerated	
		OR 2 mL serum from a serum separator (gold) tube; Minimum: 0.5 mL; Serum must be separated from cells within 48 hours of collection; Refrigerated *OR* 2 mL cerebrospinal fluid (CSF) in a sterile container; Minimum: 0.5 mL; Collectin sterile, leak-proof container; Refrigerated	
		Stability: Ambient: 72 hours Refrigerated: 28 days Frozen: 28 days	
		Methodology: Automated Nanoliter Scale Immunoassay	
		Reference Range: Refer to report	
		Days Performed: Monday–Friday	
		Reported: 4–6 days	
		СРТ: 83520 х 3	
Nickel, Urine 24 Hour	UNI24	Note: There is a clinically significant charting name change associated with this test.	8/21/17
		Includes: Creatinine, Ur mg/dL Creatinine, Ur mg/day Nickel, Urine-per volume Nickel, Urine-per 24h Nickel, Urine-ratio to CBT	
		Special Information: Collection volume MUST be indicated. Diet, medication, and nutritional supplements may introduce interfering substances. Patients should be encouraged to discontinue nutritional supplements, vitamins, minerals, and nonessential over-the-counter medications (upon the advice of their physician). High concentrations of iodine may interfere with elemental testing. Abstinence from iodine-containing medications or contrast agents for at least one month prior to collecting specimens is recommended. ARUP studies indicate that refrigeration of urine alone, during and after collection, preserves specimens adequately, if tested within 14 days of collection. Urine collected within 48 hours after administration of a gadolinium (Gd) containing contrast media (may occur with MRI studies) and acid preserved samples are unacceptable. Specimens contaminated with blood or fecal material, and specimens transported in non-trace element free transport tubes (with the exception of the original device) are also unacceptable. This test is New York DOH approved.	
		Clinical Information: Measurement of nickel is not recommended in asymptomatic individuals or in individuals with a low likelihood of exposure. Elevations in nickel urine should be interpreted with caution in individuals with no exposure risks, and may indicate contamination of the specimen.	
		Specimen Requirement: 8 mL 24-hour urine (well-mixed) in a clean container; Minimum: 1 mL; Refrigerate during collection; Collect specimen in a plastic container; Aliquot into a trace metal-free transport tube (ARUP #43116); Record total volume and collection time interval on transport tube and requisition; Refrigerated	
		OR 8 mL random urine in a clean container; Minimum: 1 mL; Refrigerate during collection; Collect specimen in a plastic container; Aliquot into a trace metal-free transport tube (ARUP #43116); Record total volume on transport tube and requisition; Refrigerated	
		(continued on page 10)	

Test Name	Order Code	Change	Effective Date
Nickel, Urine 24 Hour (continued from page 9)		Reference Range: Nickel, Urine-per volume: 0.0-10.4 μg/L Nickel, Urine-per 24h: 0.0-14.9 μg/d Nickel, Urine-ratio to CRT: 0.0-9.9 μg/g crt Creatinine, Urine mg/day: Refer to reportDays Performed: Sunday-Saturday Reported: 2-9 days	
Pancreatic Elastase, Fecal	PANCEF	Special Information: When ordering Pancreatic Elastase along with Fecal Fat, Qualitative (FFAT), please submit two separate specimens. Pancreatic Elastase needs to be sent frozen and Fecal Fat, Qualitative should be sent refrigerated. Patient Preparation: Interruption of enzyme substitution therapy recommended in order to avoid the possibility of cross-reaction with porcine enzymes. Unacceptable conditions: Stool in media or preservative; Swabs. This test is New York DOH approved. Clinical Information: Screen for exocrine pancreatic insufficiency. Normal: 201 µg/g or greater; Moderate to mild exocrine pancreatic insufficiency: 100–200 µg/g; Severe exocrine pancreatic insufficiency: 99 µg/g or less Specimen Requirement: 5 g stool in a clean container (No preservatives); Minimum: 1 g; Do not collect in media or preservative; Do not use swabs; Frozen	Effective immediately
Recombx CV2 Autoantibody Test	CV2	Special Information: Serum must be separated from cells within 48 hours of collection. Specimen Requirement: 2 mL serum from a red top tube with no additive; Minimum: 0.5 mL; Serum must be separated from cells within 48 hours of collection; Refrigerated *OR* 2 mL serum from a serum separator (gold) tube; Minimum: 0.5 mL; Serum must be separated from cells within 48 hours of collection; Refrigerated *OR* 2 mL cerebrospinal fluid (CSF) in a sterile container; Minimum: 0.5 mL; Collect in sterile, leak-proof container; Refrigerated Stability: Ambient: 72 hours Refrigerated: 28 days Frozen: 28 days Methodology: Automated Nanoliter Scale Immunoassay Reference Range: Serum: < 1:100 CSF: < 1:1 Days Performed: Monday–Friday Reported: 4–6 days CPT: 83520 x 1	8/28/17
Recombx MaTa Autoantibody Test	MATA	 Special Information: Serum must be separated from cells within 48 hours of collection. Specimen Requirement: 2 mL serum from a red top tube with no additive; Minimum: 0.5 mL; Serum must be separated from cells within 48 hours of collection; Refrigerated *OR* 2 mL serum from a serum separator (gold) tube; Minimum: 0.5 mL; Serum must be separated from cells within 48 hours of collection; Refrigerated *OR* 2 mL cerebrospinal fluid (CSF) in a sterile container; Minimum: 0.5 mL; Collect in sterile, leak-proof container; Refrigerated Stability: Ambient: 72 hours Refrigerated: 28 days Frozen: 28 days Methodology: Automated Nanoliter Scale Immunoassay Reference Range: Serum: < 1:100 CSF: < 1:1 Days Performed: Monday–Friday Reported: 4–6 days CPT: 83520 x 1	8/28/17

Test Name	Order Code	Change	Effective Date
RI Autoantibody	RIAUT	Special Information: Serum must be separated from cells within 48 hours of collection.	8/28/17
		Clinical Information: Detection of anti-Ri antibodies using recombinant human antigens.	
		Specimen Requirement: 2 mL serum from a red top tube with no additive; Minimum: 0.5 mL; Serum must be separated from cells within 48 hours of collection; Refrigerated	
		OR 2 mL serum from a serum separator (gold) tube; Minimum: 0.5 mL; Serum must be separated from cells within 48 hours of collection; Refrigerated	
		OR 2 mL cerebrospinal fluid (CSF) in a sterile container; Minimum: 0.5 mL; Collect in a sterile, leak-proof container; Refrigerated	
		Stability: Ambient: 72 hours Refrigerated: 28 days Frozen: 28 days	
		Methodology: Automated Nanoliter Scale Immunoassay Reference Range: Serum: < 1:50	
		CSF: < 1:1 Days Performed: Monday–Friday	
		Reported: 4–6 days	
		CPT: 83520 x 1	
SensoriMotor Neuropathy Profile	SENMOT	Special Information: Serum must be separated from cells within 48 hours of collection. Hemolyzed, icteric or lipemic specimens will be rejected.	8/28/17
Complete		Clinical Information: Anti-MAG, anti-SGPG, anti-GM1, anti-GD1a, anti-GD1b, anti-asialo GM1, anti-sulfatide, anti-Hu, and IgM GALOP antibodies are found in patients with a peripheral neuropathy and mixed sensory and motor symptoms. This semi-quantitative assay may be useful in monitoring therapeutic responses in select cases. Includes Co-GM1 Autoantibody Test, Co-Asialo-GM1 Autoantibody Test, Co-GD1b Autoantibody Test, GD1a Autoantibody Test, GALOP Autoantibody Test, SGPG Autoantibody Test, MAG Autoantibody Test, Recombx® Hu Autoantibody Test, and Sulfatide Autoantibody Test.	
		Specimen Requirement: 2 mL serum from a serum separator (gold) tube; Minimum: 0.5 mL; Serum must be separated from cells within 48 hours of collection; Refrigerated	
		OR 2 mL serum from a red top tube with no additive; Minimum: 0.5 mL; Serum must be separated from cells within 48 hours of collection; Refrigerated	
		Stability: Ambient: 72 hours Refrigerated: 21 days Frozen: 28 days	
		Methodology: Automated Nanoliter Scale Immunoassay Enzyme-Linked Immunosorbent Assay (ELISA) Western Blot (WB)	
		Reference Range: Refer to report	
		Days Performed: Monday, Wednesday	
		Reported: 8–11 days	
		CP1: 83520 x 11	

Test Name	Order Code	Change	Effective Date
Sensory Neuropathy Profile xp	SENNRO	Special Information: Serum must be separated from cells within 48 hours of collection. Whole blood is not an acceptable specimen. Specimens that are hemolyzed, icteric or lipemic will be rejected. Specimen Requirement: 2 mL serum from a red top tube with no additive; Minimum: 0.5 mL; Serum must be separated from cells within 48 hours of collection; Refrigerated *OR* 2 mL serum from a serum separator (gold) tube; Minimum: 0.5 mL; Serum must be separated from cells within 48 hours of collection; Refrigerated Stability: Ambient: 72 hours Refrigerated: 21 days Frozen: 28 days Methodology: Automated Nanoliter Scale Immunoassay Enzyme-Linked Immunosorbent Assay (ELISA) Western Blot (WB) Reference Range: Refer to report Days Performed: Monday, Wednesday Reported: 8–11 days CPT: 83520 x 7	8/28/17
Thallium, Blood	THALL	 Special Information: Diet, medication, and nutritional supplements may introduce interfering substances. Patients should be encouraged to discontinue nutritional supplements, vitamins, minerals, and non-essential over-the-counter medications (upon the advice of their physician). This test is New York DOH approved. Unacceptable conditions: Heparin anticoagulant, hemolyzed specimens Clinical Information: Blood thallium levels reflect recent exposure as thallium has a biological half-life of approximately two to four days. Blood levels greater than 100 µg/L are considered toxic and greater than 300 µg/L indicate severe ingestion. After severe thallium poisonings, reported symptoms have varying times of onset and include gastroenteritis, multi-organ failure and neurologic injury. Peripheral neuropathy and alopecia are well-documented effects of acute and chronic exposure. Human health effects from low-level thallium exposure are unknown. Elevated results from noncertified trace element-free collection tubes may be due to contamination. Elevated concentrations of trace elements in blood should be confirmed with a second specimen collected in a tube designed for trace element determinations, such as a royal blue (K2EDTA) or (Na2EDTA) tube. If the specimen is drawn and stored in the appropriate container, the trace element values do not change with time. Specimen Requirement: 7 mL whole blood in an EDTA royal blue top tube; Minimum: 0.5 mL; Send blood in original collection tube; Ambient Stability: Ambient: Indefinitely Refrigerated: Indefinitely Fozen: Unacceptable Days Performed: Sunday–Saturday Reported: 2–6 days 	8/21/17

Test Name	Order Code	Change	Effective Date
Thallium, Urine	UTHAL	Special Information: Patient Prep: Diet, medication, and nutritional supplements may introduce interfering substances. Patients should be encouraged to discontinue nutritional supplements, vitamins, minerals, and non-essential over-the-counter medications (upon the advice of their physician). High concentrations of iodine may interfere with elemental testing. Abstinence from iodine-containing medications or contrast agents for at least one month prior to collecting specimens for elemental testing a storemended. ARUP studies indicate that refrigeration of urine alone, during and after collection, preserves specimens adequately if tested within 14 days of collection. Record total volume and collection time interval on tube and requisition. This test is New York DOH approved. Unacceptable Conditions: Urine collected within 48 hours after administration of a gadolinium (Gd) containing contrast media (may occur with MRI studies), acid preserved urine, specimens contaminated with blood or fecal material, specimens transported in non-trace element-free transport tubes (with the exception of the original device) Clinical Information: Urinary thallium levels may reflect recent or chronic exposure, and the presence of thallium in urine after acute exposure may persist for up to several weeks. Concentrations less than 5 μ g/L are unlikely to cause adverse health effects while concentrations greater than 500 μ g/L have been associated with clinical poisoning. After severe thallium poisoning, reported symptoms have varying times of onset and include gastroenteritis, multi-organ failure and neurologic injury. Peripheral neuropative and alopecia are well-documented effects of acute and chronic exposure. Human health effects from low level thallium exposure are unknown. Specimen Requirement: 8 mL 24-hour urine (well-mixed) in a metal-free clean container; Minimum: 1 mL; Refrigerated are sellement-free transport tube (ARUP supply #43116); Record total volume on tube and requisition; Refrigerated tordinal subscontex yourse. N	8/21/17
Toluene, Blood	TOLUEN	 Special Information: Environmental/occupational exposure monitoring. Collect blood prior to the last shift of the work week. Specimens received at room temperature will be rejected. This test is New York State approved. Clinical Information: Biological Exposure Index (ACGIH): 0.02 mcg Toluene/mL blood measured in a prior to last shift of work week specimen. Reporting limit 0.3 mcg/mL Specimen Requirement: 2 mL whole blood in a potassium oxalate/sodium fluoride (gray) tube; Minimum: 0.7 mL; Refrigerated 	10/2/17
Valproic Acid, Total and Free	VPAFT2	Specimen Requirement: 3 mL plasma from a sodium or lithium heparin green top tube; Collect immediately before next dose: Refrigerated	Effective immediately
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Test Name	Order Code	Change	Effective Date
Yo Autoantibody	ANTIYO	Special Information: Serum must be separated from cells within 48 hours of collection.	8/28/17
		Clinical Information: Detection of anti-Yo antibodies using recombinant human antigens.	
		Specimen Requirement: 2 mL serum from a red top tube with no additive; Minimum: 0.5 mL; Serum must be separated from cells within 48 hours of collection; Refrigerated	
		OR 2 mL serum from a serum separator (gold) tube; Minimum: 0.5 mL; Serum must be separated from cells within 48 hours of collection; Refrigerated	
		OR 2 mL cerebrospinal fluid (CSF) in a sterile container; Minimum: 0.5 mL; Collect in a sterile, leak-proof container; Refrigerated	
		Stability: Ambient: 72 hours Refrigerated: 28 days Frozen: 28 days	
		Methodology: Automated Nanoliter Scale Immunoassay	
		Reference Range: Serum: < 1:200 CSF: < 1:1	
		Days Performed: Monday–Friday	
		Reported: 4–6 days	
		CPT: 83520 x 1	

New Tests

Test Name	Order Code	Change	Effective Date
Cytochrome P450 2D6 (CYP2D6) Geno	2D6GEN	Special Information: Unacceptable conditions: Plasma or serum; Specimens collected in sodium heparin or lithium heparin	8/21/17
2D6 (CYP2D6) Geno		 collected in sodium heparin or lithium heparin Clinical Limitation: Only the targeted CYP2D6 variants will be detected by this panel. Diagnostic errors can occur due to rare sequence variations. Risk of therapeutic failure or adverse reactions with CYP2D6 substrates may be affected by genetic and non-genetic factors that are not detected by this test. This result does not replace the need for therapeutic drug or clinical monitoring. Clinical Information: Characteristics: The cytochrome P450 (CYP) isozyme 2D6 is involved in the metabolism of many drugs, such as antiestrogens (tarnoxifen), alpha-blockers, analgesics, anticonvulsives, antidepressants, antidiabetics, antihypertensives, antipsychotics, antitusives, beta blockers, cardioactives, norepinephrine reuptake inhibitors, and stimulants. Variants of CYP2D6 will influence pharmackinetics of CYP2D6 substrates and may predict pon-standard 	
		dose requirements. Inheritance: Autosomal co-dominant. Cause: CYP2D6 gene variants and copy number result in increased, decreased or complete deficiency in enzyme activity. Variants Tested: (Variants are numbered according to M33388 sequence) Functional: *2 (2850C>T), *2A (-1584C>G; 2850C>T). Decreased function: *9 (2613-5delAGA), *10 (100C>T), *17 (1023C>T; 2850C>T), *29 (1659G>A; 2850C>T), *41 (2988G>A; 2850C>T). Non-functional: *3 (2549delA), *4 (100C>T; 1846G>A), *5 (gene deletion), *6 (1707delT), *7 (2935A>C), *8 (1758G>T; 2850C>T), *12 (124G>A; 2850C>T),	
		*14 (1758G>A; 2850C>T), *36 (a *10 carrying a CYP2D7-derived exon 9 conversion). Increased function: Duplicated functional alleles. Negative: No variants detected is predictive of *1 functional alleles and normal enzymatic activity. Allele frequencies: CYP2D6*2 or CYP2D6*2A: African-17.6%, Asian-21.2%, Caucasian-27.6%, Middle Eastern-21.7%, Oceanian-1.2%; CYP2D6*3: African-0.2%, Asian-0%, Caucasian-1.3%, Middle Eastern-0.1%, Oceanian-0.2%; CYP2D6*4: African-4.9%, Asian-4.6%, Caucasian-18.2%, Middle Eastern-7.8%,	
		Middle Eastern-2.3%, CTP2D6*0: AnhCarleo.5%, Asian-4.3%, Caticatain-2.8%, Middle Eastern-2.3%, Oceanian-4.3%; CYP2D6*6: African-0.1%, Asian-0%, Caucasian-1.0%, Middle Eastern-0.6%, Oceanian-0%; CYP2D6*8: African-0%, Caucasian-0.1%, Middle Eastern-0%, Oceanian-0%; CYP2D6*8: African-0%, Asian-0%, Caucasian-0%, Middle Eastern-0%, Oceanian-0%; CYP2D6*9: African-0.3%, Asian-0.5%, Caucasian-2.1%, Middle Eastern-0%, Oceanian-0%; CYP2D6*10: African-5.3%, Asian-30.2%, Caucasian-3.0%, Middle Eastern-3.5%, Oceanian- 2.5%; CYP2D6*12: African-0%, Asian-0%, Caucasian-0%, Middle Eastern-0%, Oceanian-0%; CYP2D6*14: African-0.1%, Asian-0%, CM2D6*17	
		Asian-0.4%, Caucasian-0%, Middle Eastern-0.2%, Oceanian-0%; CYP2D6*17: African-19.0%, Asian-0.1%, Caucasian-0.4%, Middle Eastern-1.6%, Oceanian-0.1%; CYP2D6*29: African-7.7%, Asian-0%, Caucasian-0.1%, Middle Eastern-0.8%, Oceanian-0%; CYP2D6*36: African-0.3%, Asian-0.7%, Caucasian-0%, Middle Eastern-0%, Oceanian-0%; CYP2D6*41: African-9.2%, Asian-4.9%, Caucasian-7.9%, Middle Eastern-19.9%, Oceanian-0.9%; CYP2D6xN (gene duplication): African-4.7%, Asian-1.6%, Caucasian-2.6%, Middle Eastern-7.1%, Oceanian-11.8%. Clinical Sensitivity: Drug-dependent. Analytical Sensitivity and Specificity: Greater than 99%	
		Specimen Requirement: 3 mL whole blood in an EDTA lavender top tube; Minimum: 1 mL; Refrigerated	
		OR 3 mL whole blood in an ACD A or B (yellow) tube; Minimum: 1 mL; Refrigerated	
		OR One saliva specimen in a Saliva Collection Device by Spectrum Solutions, LLC (SS-SAL-1, ARUP Supply #52535); Ambient	
		Ambient: 72 hours for whole blood; 2 weeks for saliva Refrigerated: 2 weeks for whole blood; Unacceptable for saliva Frozen: 1 month for whole blood; Unacceptable for saliva	
		Methodology: Polymerase Chain Reaction (PCR) Fluorescence Monitoring	
		Reference Range: Refer to report	
		Days Performed: Varies	
		Reported: 6-11 days	
		CPT: 81226 x 1	
		rice: \$459.00 (non-aiscountable)	

New Tests (Con't)

Test Name	Order Code	Change	Effective Date
ERBB2 (HER2/neu) Gene Amplification by FISH, Gastric Tissue	HER2GT	 Special Information: Include surgical pathology report with specimen. Preferred transport temperature is ambient. Refrigerated transport is also acceptable. Ship in cooled container during summer months. Equivocal cases will be reflexed to an alternate probe in an effort to resolve amplification status. Unacceptable conditions: Specimens fixed or processed in alternative fixatives (alcohol, Prefer) or heavy metal fixatives (B-4 or B-5). No tumor in tissue. Decalcified specimens. Tissue fixed for less than 6 hours or greater than 72 hours. This test is New York DOH approved. Clinical Information: Aid in prediction of response to HER2-directed therapy in patients with gastric cancer. Specimen Requirement: One formalin-fixed paraffin-embedded (FFPE) tissue block in a clean container; Collect tumor tissue; Formalin fix (10% neutral buffered formalin) and paraffin-embed tissue; Fixative duration: 6–72 hours; Protect paraffin block from excessive heat; Transport block in a tissue transport kit (ARUP supply #47808); Kit is recommended but not necessary; Include surgical pathology report; Ambient *OR* Five unstained FFPE slides in a clean container; Minimum: 2 slides; Collect tumor tissue; Formalin fix (10% neutral buffered formalin) and paraffin-embed tissue; Fixative duration: 6–72 hours; Protect paraffin block from excessive heat; Transport 5 unstained (4 micron thick sections) positively charged slides in a tissue transport kit (ARUP supply #47808); Kit is recommended but not necessary; Include surgical pathology report; Ambient Stability: Methodology: Fluorescent In-Situ Hybridization (FISH) Reference Range: Refer to report Days Performed: Monday–Friday Reported: 4–8 days CPT: 88377 x 1 Price: \$460,00 (non-discountable) 	8/3/17
Homocysteine, Total, Urine	UHCYS	 Special Information: Specimens received at ambient temperature will be rejected. This test is New York DOH approved. Clinical Information: To be used in conjunction with plasma amino acids and urine organic acids to aid in the biochemical screening for primary and secondary disorders of methionine metabolism. Homocysteine is an intermediary in the sulfur-amino acid metabolism pathways, linking the methionine cycle to the folate cycle. Inborn errors of metabolism that lead to homocysteinemia/-uria include cystathionine beta-synthase deficiency (homocystinuria) and various defects of methionine re-methylation. Homocysteine also was thought to be an independent predictor of cardiovascular disease (atherosclerosis, heart disease, thromboembolism), as early observational studies prior to 2000 linked homocysteine to cardiovascular risk and morbidity and mortality. However, following FDA-mandated folic acid supplementation in 1998, homocysteine for assessment of cardiovascular risk is uncertain and controversial. Based on several meta-analyses, at present, homocysteine may be regarded as a weak risk factor for coronary heart disease, and there is a lack of direct causal relationship between hyperhomocysteineina and cardiovascular disease. It is most likely an indicator of poor lifestyle and diet. Specimen Requirement: 4 mL random urine in a clean container; Minimum: 2.25 mL; Void and discard the first-morning urine specimen following an overnight fast; Continue fasting, and collect the next random urine specimen; Refrigerated Stability: Ambient: Unacceptable Refrigerated: 7 days Frozen: 2 years (continued on page 17) 	8/21/17

New Tests (Con't)

Test Name	Order Code	Change	Effective Date
Homocysteine, Total, Urine (continued from page 16)		Methodology: Liquid Chromatography-Tandem Mass Spectrometry (LC-MS/MS) Stable Isotope Dilution Reference Range: Adults: 0–9 mcmol/g creatinine Days Performed: Monday–Saturday Reported: 3–5 days CPT: 83090 x 1 Price: \$317.00 (non-discountable)	
Reticulin Antibodies, Serum	RTIABS	 Special Information: If positive, results will be titered at no additional charge. Specimens that are grossly hemolyzed or grossly lipemic will be rejected. Specimens received at ambient temperature will be rejected. This test is New York DOH approved. Clinical Information: Celiac disease (CD) is a genetically inherited autoimmune digestive disease and tends to occur in families of European descent. Family members of people with CD or dermatitis herpetiformis are at increased risk of CD. CD is characterized by a permanent intolerance to gluten. When gluten is ingested, the immune system triggers an isolated inflammatory response in the small intestinal mucosa. A lifetime gluten-free diet, the small intestine begins to repair itself and the antibody levels decline and eventually disappear. However, reintroduction of gluten-containing products stimulates the immune response again. A significant reduction in morbidity and mortality occurs when patients adhere to the gluten-free diet. Patients with CD produce various autoantibodies, including endomysial (EMA), tissue transglutaminase (TG), gliadin, and reticulin antibodies, as part of the immune response. IgA antibodies usually predominate although patients may also produce IgG autoantibodies. The levels of these antibodies decline following institution of a gluten-free diet. TG is the primary autoantigen recognized by EMA antibodies in patients with CD and is currently considered the most useful first level screening test for CD. Reticulin antibodies are no longer considered useful in the diagnosis of CD because they lack the sensitivity and specificity of the EMA and TG tests. Serological sceneling offers a minimally invasive option for rapid identification of those likely to have CD and to select those who should be subjected to biopsy. Markedly positive (serologically) individuals are highly likely to have CD and should undergo biopsy to confirm the diagnosis. Clinical Reference: 1. Murray JA: The widening spe	8/21/17

New Tests (Con't)

Test Name	Order Code	Change	Effective Date
Toxicology Screen w/ confirmation, Urine	UTOXWC	Includes: Amphetamines Barbiturates Benzodiazepines Cannabinoids Cocaine Ethanol Opiates Oxycodone Phencyclidine Special Information: Urine sent for confirmation for all components not reported	8/22/17
		Specimen Requirement: 5 mL random urine in a clean container; Minimum: 1.5 mL; Refrigerated	
		Stability: Ambient: Unacceptable Refrigerated: 5 days at 2–8 °C Frozen: Unacceptable	
		Methodology: Fluorescence Polarization Immunoassay (FPIA)	
		Reference Range: Amphetamines: Negative Barbiturates: Negative Benzodiazepines: Negative Cannabinoids: Negative Cocaine: Negative Ethanol: Negative Opiates: Negative Oxycodone: Negative Phencyclidine: Negative	
		Days Performed: 7 days per week	
		Reported: 8 hours	
		CPT: 8030/ x 1	
		File: \$78.00	

Fee Increases

Test Name	Order Code	List Fee	CPT Code	Effective Date
Lipid Associated Sialic Acid	LIPSIA	\$124.00 (non- discountable)	84275	8/21/17

Discontinued Tests

Test Name	Order Code	Test Information	Effective Date
Amniotic Bilirubin Scan	AMBILI	This test will no longer be available.	8/21/17
Bupivacaine	BUPIV	This test will no longer be available.	8/4/17
Cyto P450 2D6 Geno	2D6	This test will no longer be available. Suggest ordering Cytochrome P450 2D6 (CYP2D6) Geno (2D6GEN).	8/21/17
FISH for BK Virus		This test will no longer be available.	10/4/17
Gastric Analysis	GASTAN	This test will no longer be available.	8/21/17
Homocystine, Urine Quantitative	UHCY	This test will no longer be available. Suggest ordering Homocysteine, Total, Urine (UHCYS).	8/21/17
Leucine Aminopeptidase (LAP), Serum	LAPSER	This test will no longer be available.	8/21/17
Reticulin Antibody, IgA with reflex to Titer	RTICAB	This test will no longer be available. Suggest ordering Reticulin Antibodies, Serum (RTIABS).	8/21/17
Xylose Absorption Test (Adult)	XYLOSE	This test will no longer be available.	8/21/17
Xylose Absorption Test, Child	XYLPED	This test will no longer be available.	8/21/17