



Cleveland Clinic Laboratories

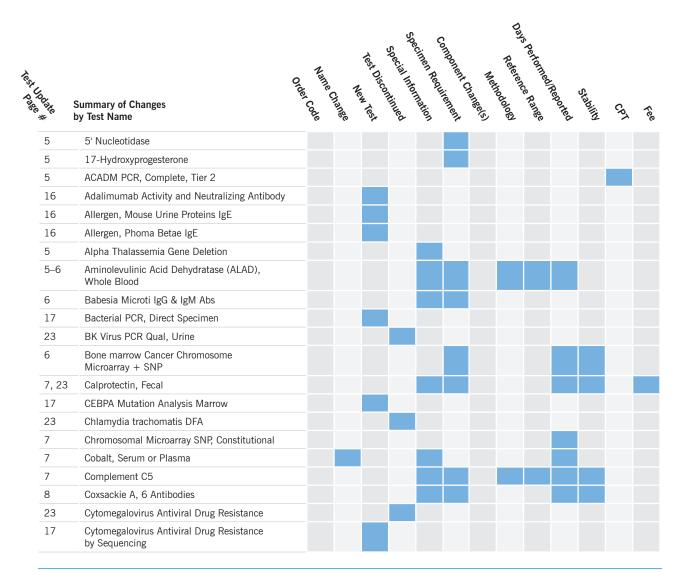
Technical Update • June 2018

Cleveland Clinic Laboratories is dedicated to keeping you updated and informed about recent testing changes. This Technical Update is provided on a monthly basis to notify you of any changes to the tests in our catalog.

Recently changed tests are bolded, and they could include revisions to methodology, reference range, days performed, or CPT code. Deleted tests and new tests are listed separately. For your convenience, tests are listed alphabetically and order codes are provided.

To compare the new information with previous test information, refer to the online Test Directory at clevelandcliniclabs. com. Test information is updated in the online Test Directory on the Effective Date stated in the Technical Update. Please update your database as necessary.

For additional detail, contact Client Services at 216.444.5755 or 800.628.6816, or via email at clientservices@ccf.org.



Summary of Changes by Test Name

Component Change (s)
Specimen Requirement
Special Information
Special Information
Reduced Code
Order Code

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18	FLT3 Tyrosine Kinase Domain Analysis Blood													
18	FLT3 Tyrosine Kinase Domain Mutation, Bone Marrow													
8	Helicobacter pylori Ab, IgG													
8	Hepatitis A Antibody, IgM													
8	Hepatitis B Core Antibody Total													
8	Hepatitis B Surface Ab, Immunity													
9	Hepatitis B Surface Ab, Qual.													
9	Hepatitis B Surface Ab, Quant													
9	Hepatitis E Antibody IgG													
9	Hepatitis E Antibody IgM													
18–19	High Sensitivity Total Testosterone													
9	HSV PCR, Miscellaneous Specimen Types													
10	Hypercoagulation Diagnostic Interpretive Panel													
10	Immunoglobulin D													
19	JAK2 Exon 12-15 Mutation Detection Bone Marrow													
19	JAK2 V617F Mutation Detection Bone Marrow													
23	Long Chain Fatty Acids													
10	Medium Chain Acyl-CoA Dehydrogenase, Tier 1, Targeted													
19	MPL Mutation Analysis Marrow													
10	MSI (PCR) X 2													
10	M. tuberculosis Amplified, CSF													
11	Mumps IgM Antibody													
11	MuSK Antibody Test													
11	Mycophenolic Acid and Metabolite													
23	Mycoplasma hominis PCR													
19	Myeloproliferative Neoplasm Panel Marrow													
19	Myeloproliferative Neoplasm Panel Peripheral Blood													
11	Neisseria meningitidis IgG Vaccine Response													
23	Norovirus Group 1 and 2 Detection by PCR													
19	NPM1 Mutation Detection Bone Marrow													
23	Oil Red O LC													
23	Oncologic CytoScan HD SNP Array													
12	Ovarian Antibody Screen with Reflex to Titer, IFA	4												
12	Phospholipids, Serum or Plasma													
12	Porphobilinogen, Urine Quant													
12	Prenatal Quad Screen													
12	Prothrombin Gene Mutation													
12	Salicylate													
13	Sickle Cell Preparation													
13	Sotalol													

Pest Nodake

Summary of Changes by Test Name

Days Performed Reported Special Information Special Information Special Information Granted Gr

23	Toxicology Screen w/ confirmation, Urine							
20	Toxicology Screen with Confirmation, Urine							
13	Tropheryma whipplei PCR							
23	Universal Bacterial, Fungal, and AFB PCR							
13	Urobilinogen Screen, Urine							
20	Urogenital Ureaplasma and Mycoplasma Species by PCR							
13	Vancomycin							
14	Vascular Endothelial Growth Factor							
21–22	Very Long-Chain and Branched-Chain Fatty Acids Profile							
14	VIP							
14	Vitamin B5 (Pantothenic Acid) Bioassay							
14	Vitamin B7 (Biotin)							
14	Vitamin C							
23	Whipple's Disease DNA by PCR							
14-15	Y-Chromosome Microdeletion							

Dear Valued Client,

For several chemistry tests, additional information regarding sample collection has been added: "Patients taking a biotin dose of up to 5 mg/day should refrain from taking biotin for 4 hours prior to sample collection. Patients taking a biotin dose of 5 to 10 mg/day should refrain from taking biotin for 8 hours prior to sample collection. Patients taking a biotin dose > 10 mg/day should consult with their physician or the laboratory prior to having a sample taken. Clinicians should consider biotin interference as a source of error, when clinically suspicious of the laboratory result." Please see the Special Information field of each affected test in the Test Directory for test-specific details. The following tests are affected:

- Beta HCG, Quantitative, Blood (HCGQT)
- Bioavailable Testosterone, SHBG, Adult Male (BTESTO)
- CA 125 (CA125)
- CA 15-3 (CA153)
- CKMB (MBE)
- CK, Total and CKMB (CKCKMB)
- DHEA-S (DHEAS)
- Estradiol-17 B (E2)
- Ferritin (FERR)
- Folate, Serum (SERFOL)
- FSH (FSH)
- LH (LH)
- LH, Pediatric (LHPED)
- LH with Tanner Stages (LHTAN)
- Luteinizing Hormone/Follicle Stimulating Hormone (XLHFSH)
- Myoglobin, Serum (MYOGLB)
- NT Pro BNP (NTBNP)
- Procalcitonin (PROCAL)

- Progesterone (PROG)
- Prolactin (PROL)
- PSA (PSA)
- PSA, Free (PSATF)
- PSA, Screening (PSAS1)
- PTH, Intact (PTHI)
- Sex Hormone Binding Globulin (SHBG2)
- T3, Free (FREET3)
- T4 (T4)
- T4, Free (FT4)
- T4/FTI (T4FTI)
- Testosterone (TESTO)
- Testosterone, Free and Total (FTESTO)
- Troponin T (TNT)
- TSH (TSH)
- Vitamin B12 (B12)
- Vitamin B12 & Folate (XB12F)
- Vitamin B12 w/reflex (B12RFX)

Test Changes

Test Name	Order Code	Change	Effective Date
5' Nucleotidase	NUC5P	Specimen Requirement: 1 mL serum from a serum separator (gold) tube; Minimum: 0.2 mL; Separate serum from cells ASAP or within 2 hours of collection; Avoid hemolysis; Refrigerated	7/31/18
17-Hydroxy- progesterone	HPROG	Specimen Requirement: 1 mL serum from a serum separator (gold) tube; Minimum: 0.3 mL (Serum, after separation from cells); Transfer 1 mL serum to standard aliquot tube; Refrigerated	Effective immediately
		OR 1 mL plasma from a sodium or lithium heparin (green) tube; Minimum: 0.3 mL (Plasma, after separation from cells); Transfer 1 mL plasma to standard aliquot tube; Refrigerated	
		OR 1 mL serum from a plain no additive (red) tube; Minimum: 0.3 mL (Serum, after separation from cells); Transfer 1 mL serum to standard aliquot tube; Refrigerated	
		OR 1 mL plasma from a sodium or lithium heparin (light green) plasma separator tube (PST); Minimum: 0.3 mL (Plasma, after separation from cells); Transfer 1 mL plasma to standard aliquot tube; Refrigerated	
ACADM PCR, Complete, Tier 2	ACADM	CPT: 81406 x 1	Effective immediately
Alpha Thalassemia Gene Deletion	ATHALS	Note: Alias names for this test include Alpha globin gene analysis, Alpha globin mutations, Alpha Thal and Alpha Thalassemia	Effective immediately
Aminolevulinic Acid Dehydratase (ALAD), Whole Blood	ALADWB	Special Information: Patient Prep: Patient should abstain from alcohol for 24 hours prior to collection. Include a list of medications the patient is currently taking. After collection, immediately place specimen in an ice bath. Hemolyzed specimens (mild to grossly hemolyzed) are unacceptable. Ambient or frozen whole blood will be rejected. Informed consent is required for New York clients. Note: Testing is not reported on Saturday or Sunday. Clinical Information: Preferred test for the confirmation of a diagnosis of aminolevulinic acid dehydratase deficiency porphyria. Aminolevulinic acid dehydratase (ALAD) activity can be inhibited in situations including hereditary tyrosinemia type 1, lead intoxication, and exposure to styrene, trichloroethylene, or bromobenzene. These causes should be ruled out when considering a diagnosis of ALAD deficiency porphyria (ADP). This method will not exhibit a decreased ALAD enzyme activity due to lead intoxication. Porphyrias are a group of inherited disorders resulting from enzyme defects in the heme biosynthetic pathway. A defect in the second enzyme of this pathway causes 5-aminolevulinic acid dehydratase deficiency porphyria. A marked deficiency of ALAD causes the accumulation and subsequent urinary excretion of large amounts of aminolevulinic acid. Urinary porphobilinogen (PBG) remains essentially normal, which rules out other forms of acute porphyria. ADP is a rare autosomal recessive acute hepatic porphyria that produces neurologic symptoms similar to those seen in acute intermittent porphyria. Symptoms include acute abdominal pain, peripheral neuropathy, nausea, vomiting, constipation, and diarrhea. Respiratory impairment, seizures, and psychosis are possible during an acute period. The workup of patients with a suspected porphyria is most effective when following a stepwise approach. Cautions: This assay is not useful in assessment of lead intoxication as it reactivates ALAD that has been inhibited by lead. Abstinence from alcohol is essential for at least 24 hours p	Effective immediately
		Specimen Requirement: 5 mL whole blood in a sodium heparin (green) tube; Minimum: 3 mL; Place specimen on ice after draw; Fill tube completely; Patient should abstain from alcohol for 24 hours; Include a list of medications the patient is currently taking; Send specimen to Cleveland Clinic Laboratories on the day of collection; Transport 5 mL whole blood in original tube; Refrigerated *OR* 5 mL whole blood in an EDTA (lavender) tube; Minimum: 3 mL; Place specimen on ice after draw; Fill tube completely; Patient should abstain from alcohol for 24 hours; Include a list of medications the patient is currently taking; Send specimen to Cleveland Clinic Laboratories on the day of collection; Transport 5 mL whole blood in original tube; Refrigerated	
		(continued on page 6)	

Test Name	Order Code	Change	Effective Date
Aminolevulinic Acid Dehydratase (ALAD), Whole Blood (continued from page 5)		*OR* 5 mL whole blood in a lithium heparin (green) tube; Minimum: 3 mL; Place specimen on ice after draw; Fill tube completely; Patient should abstain from alcohol for 24 hours; Include a list of medications the patient is currently taking; Send specimen to Cleveland Clinic Laboratories on the day of collection; Transport 5 mL whole blood in original tube; Refrigerated Methodology: Enzymatic End Point Spectrofluorometric Reference Range (Not established for patients < 16 years old): ≥ 4.0 nmol/L/sec 3.5–3.9 nmol/L/sec (Indeterminate) < 3.5 nmol/L/sec (Diminished) Days Performed: Monday, Wednesday, Friday Reported: 4–5 days	
Babesia Microti IgG & IgM Abs	BMICGM	Special Information: Bacterially contaminated, hemolyzed or lipemic specimens are unacceptable. Specimen Requirement: 1 mL serum from a serum separator (gold) tube; Minimum: 0.1 mL; Remove serum from cells ASAP or within 2 hours of collection; Parallel testing is preferred and convalescent specimens MUST be received within 30 days from receipt of the acute specimens; Please mark specimens plainly as ACUTE or CONVALESCENT; Refrigerated *OR* 1 mL serum from a plain no additive (red) tube; Minimum: 0.1 mL; Remove serum from cells ASAP or within 2 hours of collection; Parallel testing is preferred and convalescent specimens MUST be received within 30 days from receipt of the acute specimens; Please mark specimens plainly as ACUTE or CONVALESCENT; Refrigerated *OR* 1 mL plasma from an EDTA (lavender) tube; Minimum: 0.1 mL; Remove plasma from cells ASAP or within 2 hours of collection; Parallel testing is preferred and convalescent specimens MUST be received within 30 days from receipt of the acute specimens; Please mark specimens plainly as ACUTE or CONVALESCENT; Refrigerated *OR* 1 mL plasma from a sodium or lithium heparin (green) tube; Minimum: 0.1 mL; Remove plasma from cells ASAP or within 2 hours of collection; Parallel testing is preferred and convalescent specimens MUST be received within 30 days from receipt of the acute specimens; Please mark specimens plainly as ACUTE or CONVALESCENT; Refrigerated *OR* 1 mL plasma from an ACD A or B (yellow) tube; Minimum: 0.1 mL; Remove plasma from cells ASAP or within 2 hours of collection; Parallel testing is preferred and convalescent specimens MUST be received within 30 days from receipt of the acute specimens MUST be received within 30 days from receipt of the acute specimens MUST be received within 30 days from receipt of the acute specimens; Please mark specimens plainly as ACUTE or CONVALESCENT; Refrigerated	Effective immediately
Bone marrow Cancer Chromosome Microarray + SNP	BMHSNP	Specimen Requirement: 4 mL bone marrow in an EDTA (lavender) tube; Notes: Collect specimen Monday–Friday only; If aliquoting is necessary, sterile aliquot tubes must be used; Ambient *OR* 5–7 mL bone marrow in a sodium heparin (green) tube; EDTA is preferred container type; Collect specimen Monday–Friday only; If aliquoting is necessary, sterile aliquot tubes must be used; Ambient Stability: Ambient: 48 hours Refrigerated: Not preferred Frozen: Unacceptable Days Performed: Monday–Friday Reported: 14 days	6/26/18

Test Name	Order Code	Change	Effective Date
Calprotectin, Fecal	CALPRO	Note: Clinical Information will be removed for this test Specimen Requirement: 5 g random stool in a sterile container; Minimum: 1 g; Collect stool in a preservative-free, sterile container; Refrigerated Stability: Ambient: 10 days Refrigerated: 10 days Frozen: 1 year Days Performed: Monday, Wednesday, Friday Reported: 2–4 days	7/31/18
Chromosomal Microarray SNP, Constitutional	CRMSNP	Days Performed: 3 days per week Reported: 14 days	6/7/18
Cobalt, Serum or Plasma	COBALT	Test Name: Previously Cobalt, Serum Special Information: Patient Prep: Diet, medication and nutritional supplements may introduce interfering substances. Patients should be encouraged to discontinue nutritional supplements, vitamins, minerals and non-essential over-the-counter medications (upon the advice of their physician). Specimens that are not separated from the red cells, or clot, within 6 hours are unacceptable. Separator tubes will be rejected. This test is New York DOH approved.	Effective immediately
		Clinical Information: Occupational exposure or toxic ingestion monitoring. Preferred test for evaluating metal ion release from metal-on-metal joint arthroplasty. Levels may be increased in asymptomatic patients with metal-on-metal prosthetics and should be considered in the context of the overall clinical scenario. Symptoms associated with cobalt toxicity vary based on route of exposure, and may include cardiomyopathy, allergic dermatitis, pulmonary fibrosis, cough and dyspnea. Elevated results from noncertified trace element-free tubes may be due to contamination. Elevated concentrations of trace elements in serum should be confirmed with a second specimen collected in a navy blue top tube with no additive.	
		Days Performed: Sunday–Saturday Reported: 2–6 days	
Complement C5	COMPC5	Special Information: CRITICAL FROZEN. Separate specimens must be submitted when multiple tests are ordered. Non-frozen specimens will be rejected. Specimens left to clot at refrigerated temperature and specimens exposed to repeated freeze/thaw cycles are unacceptable. This test is New York DOH approved. Clinical Information: Used as a follow-up test for complement activity screening when CH50 and AH50 are low or absent and high suspicion remains for complement deficiency.	8/7/18
		Specimen Requirement: 1 mL serum from a serum separator (gold) tube; Minimum: 0.3 mL; Allow specimen to clot for one hour at room temperature; Separate serum from cells ASAP or within 2 hours of collection and transfer into standard aliquot tube; Freeze immediately; Separate specimens must be submitted when multiple tests are ordered; Critical Frozen Stability: Ambient: After separation from cells: 2 hours Refrigerated: After separation from cells: Unacceptable Frozen: After separation from cells: 2 weeks (Avoid repeated freeze/thaw cycles)	
		Methodology: Radial Immunodiffusion (RID) Reference Range: 7–20 mg/dL Days Performed: Tuesday, Friday Reported: 4–9 days	

Test Name	Order Code	Change	Effective Date
Coxsackie A, 6 Antibodies	COXSA6	Special Information: This test is New York DOH approved. Specimen Requirement: 2 mL serum from a plain no additive (red) tube; Minimum: 1 mL; Centrifuge and transfer serum to a standard aliquot tube; Ambient *OR* 2 mL serum from a serum separator (gold) tube; Minimum: 1 mL; Centrifuge and transfer serum to a standard aliquot tube; Ambient Stability: Ambient: 1 week Refrigerated: 2 weeks Frozen: 1 month Days Performed: Varies Reported: 4–9 days	8/9/18
Helicobacter pylori Ab, IgG	HPYLRI	Days Performed: Monday–Friday Reported: 1–3 days	Effective immediately
Hepatitis A Antibody, IgM	AHAVM	Special Information: Not intended for use in screening blood, plasma, or tissue donors. Performance has not been established for the use of cadaveric specimens or the use of body fluids other than human serum. A reactive IgM anti-HAV result does not necessarily rule out other hepatitis infections. The results should be used and interpreted only in the context of the overall clinical picture. A negative test result does not exclude the possibility of exposure to hepatitis A virus. Specimens from individuals with Non-Hodgkin's Lymphoma may cross-react with this assay. Clinical Limitation: Criteria for rejection: Heat inactivated, pooled, grossly hemolyzed, obvious microbial contamination	6/12/18
Hepatitis B Core Antibody Total	AHBCOT	Special Information: Not intended for use in screening blood, plasma, or tissue donors. Assay performance characteristics have not been established for immunocompromised or immunosuppressed patients. Performance has not been established for the use of cadaveric specimens or the use of body fluids other than human serum. A nonreactive test result does not exclude the possibility of exposure to or infection with Hepatitis B Virus (HBV). Clinical Limitation: Criteria for rejection: Heat inactivated, pooled, grossly hemolyzed, obvious microbial contamination	6/12/18
Hepatitis B Surface Ab, Immunity	AHBSI	Special Information: For diagnostic purposes, results should be used in conjunction with patient history and other hepatitis markers for diagnosis of acute and chronic infection. Performance characteristics have not been established for therapeutic monitoring. Test is not intended for use in screening blood, plasma, or tissue donors. Performance characteristics have not been established for immunocompromised or immunosuppressed patients; results should be interpreted with caution. Performance has not been established for the use of cadaveric specimens. A non-reactive test result does not exclude the possibility of exposure to hepatitis B virus (HBV). Assay does not differentiate between vaccines and natural infections. A reactive anti-HBs result does not exclude co-infection by another hepatitis virus. Results obtained with the ARCHITECT AUSAB assay may not be used interchangeably with values obtained with different manufacturers' assay methods. Clinical Information: To assess the presence of a recent or remote immune response to HBV infection or vaccination. Specimen Requirement: 0.5 mL serum from a serum separator (gold) tube; Minimum: 0.25 mL; Refrigerated *OR* 0.5 mL plasma from an EDTA (lavender) tube; Minimum: 0.25 mL; Refrigerated	6/4/18

Test Name	Order Code	Change	Effective Date
Hepatitis B Surface Ab, Qual.	AHBSAG	Special Information: For diagnostic purposes, results should be used in conjunction with patient history and other hepatitis markers for diagnosis of acute and chronic infection. Performance characteristics have not been established for therapeutic monitoring. Test is not intended for use in screening blood, plasma, or tissue donors. Performance characteristics have not been established for immunocompromised or immunosuppressed patients; results should be interpreted with caution. Performance has not been established for the use of cadaveric specimens. A non-reactive test result does not exclude the possibility of exposure to hepatitis B virus (HBV). Assay does not differentiate between vaccines and natural infections. A reactive anti-HBs result does not exclude co-infection by another hepatitis virus. Results obtained with the ARCHITECT AUSAB assay may not be used interchangeably with values obtained with different manufacturers' assay methods. Clinical Limitation: Criteria for Rejection: Heat-inactivated, pooled, grossly hemolyzed, obvious microbial contamination Clinical Information: To assess adequacy of recent or remote immune response to HBV infection or vaccination. Specimen Requirement: 0.5 mL serum from a serum separator (gold) tube; Minimum: 0.25 mL; Refrigerated *OR* 0.5 mL plasma from an EDTA (lavender) tube; Minimum: 0.25 mL; Refrigerated	6/4/18
Hepatitis B Surface Ab, Quant	AHBSQ	Special Information: For diagnostic purposes, results should be used in conjunction with patient history and other hepatitis markers for diagnosis of acute and chronic infection. Performance characteristics have not been established for therapeutic monitoring. Test is not intended for use in screening blood, plasma, or tissue donors. Performance characteristics have not been established for immunocompromised or immunosuppressed patients; results should be interpreted with caution. Performance has not been established for the use of cadaveric specimens. A non-reactive test result does not exclude the possibility of exposure to hepatitis B virus (HBV). Assay does not differentiate between vaccines and natural infections. A reactive anti-HBs result does not exclude co-infection by another hepatitis virus. Results obtained with the ARCHITECT AUSAB assay may not be used interchangeably with values obtained with different manufacturers' assay methods. Clinical Information: To assess the presence of a recent or remote immune response	6/4/18
		to HBV infection or vaccination. Specimen Requirement: 0.5 mL serum from a serum separator (gold) tube; Minimum: 0.25 mL; Refrigerated *OR* 0.5 mL plasma from an EDTA (lavender) tube; Minimum: 0.25 mL; Refrigerated Stability: Ambient: 3 days Refrigerated: 7 days Frozen: 14 days	
Hepatitis E Antibody IgG	HEPIGG	Days Performed: Tuesday, Thursday, Saturday Reported: 2–6 days	Effective immediately
Hepatitis E Antibody IgM	HEPIGM	Days Performed: Tuesday, Thursday, Saturday Reported: 2–6 days	Effective immediately
HSV PCR, Miscellaneous Specimen Types	PCRHSV	Stability: Ambient: Tissue: Unacceptable; Plasma, serum, amniotic fluid, BAL: 8 hours Refrigerated: Tissue: Unacceptable; Plasma, serum, amniotic fluid, BAL: 72 hours Frozen: 3 months	Effective immediately

Test Name	Order Code	Change	Effective Date
Hypercoagulation Diagnostic Interpretive Panel	HYPER	Specimen Requirement: 2 mL serum from a serum separator (gold) tube; Minimum: 1 mL; Indicate clearly which tube is plasma and which tube is serum; Submit Coagulation Consultation Patient History Form; Frozen *AND* 2 mL plasma from an EDTA (lavender) tube; Minimum: 1 mL; Refrigerated *AND* 4 mL whole blood in an EDTA (lavender) tube; Minimum: 2 mL; Refrigerated *AND* 6 mL plasma from a 3.2% sodium citrate (light blue) tube; Minimum: 3 mL; Frozen Reference Range: Refer to individual components (See Homocysteine)	6/7/18
Immunoglobulin D	IGDQNT	Special Information: Contaminated, hemolyzed or severely lipemic specimens are unacceptable. This test is New York DOH approved. Specimen Requirement: 1 mL serum from a serum separator (gold) tube; Minimum: 0.5 mL; Separate serum from cells ASAP or within 2 hours of collection and transfer into standard aliquot tube; Frozen Stability: Ambient: After separation from cells: 8 hours Refrigerated: After separation from cells: 48 hours Frozen: After separation from cells: 1 month	7/31/18
Medium Chain Acyl- CoA Dehydrogenase, Tier 1, Targeted	MCADD	Methodology: Sanger Sequencing CPT: 81403 x 1	Effective immediately
MSI (PCR) X 2	MSICCT	Specimen Requirement: Representative tumor: Formalin-fixed, paraffin-embedded (FFPE) tissue block, OR 10 unstained slides *AND* Representative patient normal: Formalin-fixed, paraffin-embedded (FFPE) tissue block, 5 unstained slides, preferably from separate block *OR* Representative tumor: Formalin-fixed, paraffin-embedded (FFPE) tissue block, OR 10 unstained slides *AND* 4 mL whole blood in an EDTA (lavender) tube; Representative patient normal: Whole blood if no FFPE tissue is available (e.g., small biopsies); Ambient Stability: Ambient: Whole Blood–24 hours; Paraffin–embedded tissue–Indefinitely Refrigerated: Whole blood–7 days Frozen: Unacceptable Days Performed: 2 days per week Reported: 7 days	6/26/18
M. tuberculosis Amplified, CSF	MTBCSF	For Interfaced Clients Only: Test build may need to be modified Special Information: Refer to specimen requirements for client-processed specimens. Blood, urine, swabs, stool, paraffin blocks and tissue are unacceptable. This test is New York DOH approved. Specimen source is required. In order to perform testing, it is essential to know whether or not the submitted specimen has been processed (digestion and decontamination procedure). If processed, smear results must be provided as a comment on the test order or requisition. Turnaround time will be delayed if the required information is not provided. Clinical Information: This panel uses PCR testing to detect M. tuberculosis complex isolates and determine possible resistance to rifampin treatment. Specimen Requirement: 5–10 mL cerebrospinal fluid (CSF) in a sterile container; Minimum: 1 mL; Label as 'Unprocessed CSF;' Specimen source required; Refrigerated *OR* 2–5 mL digested/decontaminated cerebrospinal fluid (CSF) in a sterile container; Minimum: 1 mL; Indicate as "Processed CSF' and provide smear results and identify method of digestion; Each specimen should be placed in an individually sealed bag; Refrigerated Stability: Ambient: Unprocessed: 3 days; Processed: Unacceptable Refrigerated: Unprocessed: 1 week; Processed: 1 week Frozen: Unprocessed: 1 month; Processed: 1 month Methodology: Qualitative Polymerase Chain Reaction CPT: 87556 x 1, 87798 x 1	8/7/18

Test Name	Order Code	Change	Effective Date
Mumps IgM Antibody	MUMPSM	Special Information: Acute and convalescent specimens must be labeled as such; parallel testing is preferred and convalescent specimens MUST be received within 30 days from receipt of the acute specimens. Please mark specimen plainly as 'acute' or 'convalescent.' Contaminated, hemolyzed, heat-inactivated, or severely lipemic specimens are unacceptable. Specimen Requirement: 1 mL serum from a serum separator (gold) tube; Minimum: 0.2 mL; Separate serum from cells ASAP or within 2 hours of collection; Refrigerated Stability: Ambient: 2 days Refrigerated: 14 days Frozen: 1 year (Avoid repeated freeze/thaw cycles) Days Performed: Monday–Friday Reported: 2–6 days	Effective immediately
MuSK Antibody Test	MUSK	Special Information: This test is New York State approved. Clinical Information: Testing is useful for: 1. Diagnosis of autoimmune muscle-specific kinase (MuSK) myasthenia gravis (MG) 2. Second-line test aiding diagnosis of autoimmune MG in patients with new onset acquired MG evident clinically and electrophysiologically, but negative first-line serological tests 3. Establishing a quantitative baseline value for MuSK antibodies that allows comparison with future levels if weakness is worsening. A positive result, in the appropriate clinical context, confirms the diagnosis of autoimmune muscle-specific kinase MG. Seropositivity justifies consideration of immunotherapy. Cautions: Immunosuppressant therapy is a common cause of false-seronegativity. It is, therefore, important to perform a comprehensive serological evaluation before initiating immunosuppressant therapy. Interpretation of a patient's serological and clinical status is further complicated when characteristic signs of MG are obscured by a superimposed steroid-induced myopathy. Specimen Requirement: 1.5 mL serum from a plain no additive (red) tube; Minimum: 1 mL; Refrigerated *OR* 1.5 mL serum from a serum separator (gold) tube; Minimum: 1 mL; Refrigerated *CR* 1.5 mL serum from a serum separator (gold) tube; Minimum: 1 mL; Refrigerated: 28 days Frozen: 28 days Reference Range: ≤ 0.02 nmol/L Days Performed: Tuesday, Thursday Reported: 4—11 days	8/2/18
Mycophenolic Acid and Metabolite	MYCMET	Clinical Information: Therapeutic monitoring for individuals taking mycophenolate. The therapeutic range is based on serum pre-dose (trough) draw at steady state concentration. A proposed therapeutic range is 1.0–3.5 μg/mL for a 2 g/day dose. A 3 g/day dose may have plasma concentrations up to 5.0 μg/mL. Trough concentrations between 2.0–4.0 μg/mL have been suggested to maximize efficacy and minimize adverse effects. Mycophenolic acid glucuronide is an inactive metabolite and a range of 35.0–100.0 μg/mL indicates normal metabolism. During the first two weeks of transplantation, mycophenolic acid glucuronide concentrations are typically 100–250 μg/mL. Adverse effects of toxicity include abdominal pain, peripheral edema, cardiac abnormalities, hypertension and electrolyte disturbances. Reference Range: Mycophenolic Acid 0–99 Years: 1.0–3.5 μg/mL Myc. Acid Glucuronide 0–99 Years: 35.0–100.0 μg/mL	Effective immediately
Neisseria meningitidis IgG Vaccine Response	NMEN	Days Performed: Tuesday Reported: 2–8 days	Effective immediately

Test Name	Order Code	Change	Effective Date
Test Name Ovarian Antibody Screen with Reflex to Titer, IFA	Ovaran	Change For Interfaced Clients Only: Test build may need to be modified Test Name: Previously Ovarian Antibody Special Information: If Ovarian Antibody Screen is positive, a titer will be performed at an additional cost. Grossly hemolyzed and grossly lipemic specimens will be rejected. Clinical Information: Ovarian Antibody is found in patients with premature ovarian failure, Addison's disease, and polyendocrinopathy syndromes. Specimen Requirement: 1 mL serum from a plain no additive (red) tube; Minimum: 0.3 mL; Do not use serum separator tubes; Send using cold packs; Refrigerated Stability: Ambient: 7 days Refrigerated: 14 days Frozen: 30 days Methodology: Immunofluorescence Reference Range: Negative; Ovarian antibody screen is at 1:5 dilution Days Performed: Wednesday Reported: 4–11 days CPT: 86255 x 1	Effective Date 6/7/18
Phospholipids, Serum or Plasma	PHOLIP	Test Name: Previously Phospholipids, Serum Special Information: Patient Prep: Patient should fast for 12 hours prior to specimen collection. This test is New York DOH approved. Clinical Information: This test is useful in the evaluation of liver disease, particularly obstructive jaundice. Specimen Requirement: 1 mL serum from a serum separator (gold) tube; Minimum: 0.5 mL; Collect blood following a 12-hour fast; Allow specimen to clot completely at room temperature, then transfer serum into a standard aliquot tube; Refrigerated *OR* 1 mL plasma from an EDTA (lavender) tube; Minimum: 0.5 mL; Collect blood following a 12-hour fast; Allow specimen to clot completely at room temperature, then transfer plasma into a standard aliquot tube; Refrigerated *OR* 1 mL plasma from a sodium or lithium heparin (green) tube; Minimum: 0.5 mL; Collect blood following a 12-hour fast; Allow specimen to clot completely at room temperature, then transfer plasma into a standard aliquot tube; Refrigerated Stability: Ambient: After separation from cells: 8 hours Refrigerated: After separation from cells: 1 month Frozen: After separation from cells: 1 month Methodology: Spectrophotometry Reference Range: 160–300 mg/dL Days Performed: Monday, Wednesday, Friday Reported: 2–5 days	8/2/18
Porphobilinogen, Urine Quant	UPBGQT	Special Information: Indicate total volume and time of collection on requisition. Body fluids other than urine are unacceptable. Specimen Requirement: 8 mL 24-hour urine (well-mixed) in a clean container; Minimum: 3.5 mL; Refrigerate during collection; Protect specimen from light; Frozen *OR* 8 mL random urine in a clean container; Minimum: 3.5 mL; Protect specimen from light; Frozen	Effective immediately
Prenatal Quad Screen	QUAD4	Stability: Ambient: 1 day Refrigerated: 7 days Frozen: 14 days	7/31/18
Prothrombin Gene Mutation	PTGEN	Days Performed: 3 days per week Reported: 5 days	Effective immediately

Test Name	Order Code	Change	Effective Date
Salicylate	SALI	Reference Range: 0-99 Years: 3.0-30.0 mg/dL (See Clinical Information)	Effective immediately
Sickle Cell Preparation	SCKSOL	Special Information: If the Sickle Cell Preparation is positive, the further evaluation by capillary electrophoresis will be performed at an additional charge. The Sickle Cell Preparation test should not be performed on children less than 6 months old. For children less than 6 months of age, the hemoglobin evaluation cascade (HBEVAL) should be ordered. Further testing may be required which includes high performance liquid chromatography, isoelectric focusing, and rarely alkaline and acid electrophoresis. These tests will be performed at an additional charge.	Effective immediately
Sotalol	SOTAL	Special Information: Draw specimen prior to next dose—at steady state concentration. Please provide the following information if available: 1.) Dose—List drug amount and include the units of measure 2.) Route—List the route of administration (IV, oral, etc.) 3.) Dose Frequency—Indicate how often the dose is administered (per day, per week, as needed, etc.) 4.) Type of Draw—Indicate the type of blood draw (Peak, trough, random, etc.) Whole blood is unacceptable. Gel separator tubes, light blue (sodium citrate), and yellow (SPS or ACD solution) tubes will be rejected. Specimen Requirement: 1 mL serum from a plain no additive (red) tube; Minimum: 0.5 mL; Pre-dose (trough) draw—At a steady state concentration; Separate serum from cells within 2 hours of collection; Refrigerated *OR* 1 mL plasma from an EDTA (lavender) tube; Minimum: 0.5 mL; Pre-dose (trough) draw—At a steady state concentration; Separate plasma from cells within 2 hours of collection; Refrigerated Days Performed: Monday, Thursday Reported: 2—6 days	Effective immediately
Tropheryma whipplei PCR	WHIPWB	Special Information: Must indicate specimen source. Heparinized specimens are not acceptable. This test is New York DOH approved. Specimen Requirement: 1 mL whole blood from an EDTA (lavender) tube; Minimum: 0.5 mL; Transfer 1 mL whole blood into sterile aliquot tube; Specimen source required; Frozen *OR* 1 mL plasma from an EDTA (lavender) tube; Minimum: 0.5 mL; Transfer 1 mL plasma into sterile aliquot tube; Specimen source required; Frozen *OR* 1 mL serum from a serum separator (gold) tube; Minimum: 0.5 mL; Transfer 1 mL serum into sterile aliquot tube; Specimen source required; Frozen *OR* 1 mL cerebrospinal fluid (CSF) in a sterile container; Minimum: 0.5 mL; Specimen source required; Frozen *OR* Tissue in a sterile container; Freeze immediately; Specimen source required; Frozen *OR* Formalin-fixed paraffin-embedded (FFPE) tissue in a sterile container; Specimen source required; Ambient Stability: Ambient: Whole blood, plasma, serum, CSF: 24 hours; Tissue: Unacceptable; FFPE: Indefinitely Refrigerated: Whole blood, plasma, serum, CSF: 1 month; Tissue: 1 month; FFPE: Unacceptable	7/31/18
Urobilinogen Screen, Urine	UUROB	Stability: Ambient: Clean container–2 hours / BD Urine Preservative tube–72 hours Refrigerated: 24 hours	Effective immediately

Test Name	Order Code	Change	Effective Date
Vancomycin	VANCRA	Specimen Requirement: 1 mL serum from a plain no additive (red) tube; Minimum: 0.5 mL; Do not collect in a gel separator tube; Centrifuge and transfer serum to a CCL tube and refrigerate; Refrigerated *OR* 1 mL plasma from an EDTA (lavender) tube; Minimum: 0.5 mL; Do not collect in a gel separator tube; Centrifuge and transfer plasma to a CCL tube and refrigerate; Refrigerated *OR* 1 mL plasma from a lithium heparin (green) tube; Minimum: 0.5 mL; Do not collect in a gel separator tube; Centrifuge and transfer plasma to a CCL tube and refrigerate; Refrigerated	Effective immediately
Vascular Endothelial Growth Factor	VEGF	Special Information: This assay is performed using the QuantiGlo® Chemiluminescent EIA kit. Values obtained with different assay methods or kits cannot be used interchangeably. Hemolyzed specimens are unacceptable.	Effective immediately
VIP	VIP	Special Information: Grossly hemolyzed specimens are unacceptable. Peripheral draws are preferred. Samples drawn (directly into the protease tube) from ports are acceptable, but not recommended, and run the risk of contamination. Separate specimens must be submitted when multiple tests are ordered. This test is New York DOH approved.	Effective immediately
Vitamin B5 (Pantothenic Acid) Bioassay	VITB5	Special Information: Grossly hemolyzed, warm greater than 24 hours, grossly lipemic, specimens containing particulate matter or microbial contamination, and specimens unprotected from light are unacceptable. Specimen Requirement: 1 mL serum from a serum separator (gold) tube; Minimum: 0.5 mL; Aliquot serum into a plastic tube and protect from light; Frozen Stability: Ambient: Unacceptable Refrigerated: 7 days Frozen: 21 days Days Performed: Monday Reported: 3–13 days	7/31/18
Vitamin B7 (Biotin)	VITB7	Special Information: Grossly hemolyzed, grossly lipemic, thawed (warm or cold), and specimens not light protected are unacceptable. Days Performed: Monday Reported: 2–18 days	Effective immediately
Vitamin C	VITC	Special Information: Thawing and refreezing of the specimen and exposure to light will result in decreased Vitamin C concentration. EDTA plasma, whole blood, body fluids, and grossly hemolyzed specimens are unacceptable. Stability: Ambient: After separation from cells: Unacceptable Refrigerated: After separation from cells: Unacceptable Frozen: After separation from cells: 1 year Days Performed: 2–7 days Reported: Sunday, Tuesday–Thursday, Saturday	Effective immediately

Test Name	Order Code	Change	Effective Date
Y-Chromosome Microdeletion	YCMICR	For Interfaced Clients Only: Test build may need to be modified Special Information: Counseling and informed consent are recommended for genetic testing. Do NOT freeze. Serum, frozen specimens, and severely hemolyzed specimens are unacceptable. This test is New York DOH approved. Clinical Limitation: Diagnostic errors can occur due to rare sequence variations. Mutations within individual genes included in the AZF regions will not be detected. Breakpoints of identified microdeletions will not be determined. Male infertility, due to causes other than Y chromosome microdeletions tested, has not been excluded. Clinical Information: Aids in determining the cause of azoospermia or oligospermia and helps predict effectiveness of assisted reproductive technologies in men with Y chromosome microdeletions. Background Information: Y Chromosome Microdeletion. Characteristics: Y chromosome microdeletions are typically characterized by azoospermia, severe to moderate oligospermia, or abnormal sperm morphology/motility in men with a normal physical evaluation. Assisted reproductive techniques are contraindicated for men carrying AZFa, AZFb, AZFbc or AZFabc microdeletions, which are classically associated with spermatogenic failure. Prevalence: 1 in 2,000 to 3,000 males carry Y chromosome deletions/microdeletions. Penetrance: Approaches 100% in males; variable expression may result in intra-familial variation of fertility in men with an identical microdeletion. Inheritance: Y-linked; microdeletions are typically de novo. Cause: Microdeletion. Inheritance: Y-linked; microdeletions are typically de novo. Cause: Microdeletions of the Y chromosome azoospermia factor regions a, b or c (AZFa, AZFb or AZFc). Mutations Tested: Five common Y chromosome microdeletions: AZFa, AZFb, AZFbc, and AZFabc.	8/7/18
Y-Chromosome Microdeletion		Clinical Sensitivity: Estimated at 5 to 10% for men with non-obstructive azoospermia or severe oligospermia. Methodology: Multiplex PCR followed by	
(continued from page 14)		electrophoresis. Analytical Sensitivity and Specificity: Greater than 99% Specimen Requirement: 3 mL whole blood in an EDTA (lavender) tube; Minimum: 1 mL; Do NOT freeze; Send to Cleveland Clinic Laboratories on the day of collection; Refrigerated	

Y-Chromosome Microdeletion (continued from page 14)	Clinical Sensitivity: Estimated at 5 to 10% for men with non-obstructive azoospermia or severe oligospermia. Methodology: Multiplex PCR followed by electrophoresis. Analytical Sensitivity and Specificity: Greater than 99% Specimen Requirement: 3 mL whole blood in an EDTA (lavender) tube; Minimum: 1 mL; Do NOT freeze; Send to Cleveland Clinic Laboratories on the day of collection; Refrigerated	
	OR 3 mL whole blood in a sodium or lithium heparin (green) tube; Minimum: 1 mL; Do NOT freeze; Send to Cleveland Clinic Laboratories on the day of collection; Refrigerated	
	OR 3 mL whole blood in a sodium citrate (light blue) tube; Minimum: 1 mL; Do NOT freeze; Send to Cleveland Clinic Laboratories on the day of collection; Refrigerated	
	OR 3 mL whole blood in an ACD A or B (yellow) tube; Minimum: 1 mL; Do NOT freeze; Send to Cleveland Clinic Laboratories on the day of collection; Refrigerated	
	Stability: Ambient: 72 hours Refrigerated: 1 week Frozen: Unacceptable	
	Methodology: Electrophoresis Polymerase Chain Reaction (PCR)	
	Days Performed: Tuesday, Friday	
	Reported: 8–11 days	

New Tests

Test Name	Order Code	Change	Effective Date
Adalimumab Activity and Neutralizing Antibody	ADANEU	Note: This test was previously announced in the May 2018 Technical Update. Price: \$389.00 (non-discountable)	Effective immediately
Allergen, Mouse Urine Proteins IgE	MOUUR	Clinical Information: Specific evaluation of allergic reactions. IgE (kU/L) Interpretation: < 0.35, Class 0–Below Detection; 0.35–0.69, Class 1–Low; 0.70–3.49, Class 2–Moderate; 3.50–17.49, Class 3–High; 17.50–49.99, Class 4–Very High; 50.00–99.99, Class 5–Very High; ≥ 100, Class 6–Very High Specimen Requirement: 0.5 mL serum from a serum separator (gold) tube; Minimum: 0.3 mL (Submitting the minimum volume will not allow for repeat testing or add-ons. Required volume of 0.5 mL is preferred when possible); An extra 50 μ L will be required for each additional allergen ordered; Refrigerated *0R* 0.5 mL plasma from a lithium heparin (light green) plasma separator tube (PST); Minimum: 0.3 mL (Submitting the minimum volume will not allow for repeat testing or add-ons. Required volume of 0.5 mL is preferred when possible); An extra 50 μ L will be required for each additional allergen ordered; Refrigerated *0R* 0.5 mL plasma from an EDTA (lavender) tube; Minimum: 0.3 mL (Submitting the minimum volume will not allow for repeat testing or add-ons. Required volume of 0.5 mL is preferred when possible); An extra 50 μ L will be required for each additional allergen ordered; Refrigerated Stability: Ambient: 1 day Refrigerated: 7 days Frozen: 14 days Methodology: Fluorescent Enzyme Immunoassay (FEIA) by ImmunoCAP Reference Range: Allergen, Mouse Urine Proteins IgE: < 0.35 kU/L Allergen, Mouse Urine Proteins Class: 0 Days Performed: Sunday–Saturday Reported: 1–2 days CPT: 86003 x 1	7/31/18
Allergen, Phoma Betae IgE	PHOMAB	Clinical Information: Specific evaluation of allergic reactions. IgE (kU/L) Interpretation: < 0.35, Class 0–Below Detection; 0.35–0.69, Class 1–Low; 0.70–3.49, Class 2–Moderate; 3.50–17.49, Class 3–High; 17.50–49.99, Class 4–Very High; 50.00–99.99, Class 5–Very High; \geq 100, Class 6–Very High Specimen Requirement: 0.5 mL serum from a serum separator (gold) tube; Minimum: 0.3 mL (Submitting the minimum volume will not allow for repeat testing or add-ons. Required volume of 0.5 mL is preferred when possible); An extra 50 μ L will be required for each additional allergen ordered; Refrigerated *OR* 0.5 mL plasma from an EDTA (lavender) tube; Minimum: 0.3 mL (Submitting the minimum volume will not allow for repeat testing or add-ons. Required volume of 0.5 mL is preferred when possible); An extra 50 μ L will be required for each additional allergen ordered; Refrigerated *OR* 0.5 mL plasma from a lithium heparin (light green) plasma separator tube (PST); Minimum: 0.3 mL (Submitting the minimum volume will not allow for repeat testing or add-ons. Required volume of 0.5 mL is preferred when possible); An extra 50 μ L will be required for each additional allergen ordered; Refrigerated Stability: Ambient: 1 day Refrigerated: 7 days Frozen: 14 days Methodology: Fluorescent Enzyme Immunoassay (FEIA) by ImmunoCAP Reference Range: Allergen, Phoma Betae IgE: < 0.35 kU/L Allergen, Phoma Betae Class: 0 Days Performed: Sunday–Saturday Reported: 1–2 days CPT: 86003 x 1	7/31/18

Test Name	Order Code	Change	Effective Date
Bacterial PCR, Direct Specimen	BCTPCR	Special Information: Rejection Criteria: Swabs will be rejected. Test will not be performed on blood or blood components. Specimens from non-sterile sites will be rejected. Specimens received in formalin or other transport media will be rejected. Clinical Limitation: This test will not be able to identify organisms in a polymicrobial population. This test is not validated for sterile fluids or fresh-frozen plasma-embedded (FFPE) samples. These samples will be sent to a reference laboratory for testing. Clinical Information: Routine culture results for orders will be reviewed to determine if testing is indicated. Specimen Requirement: One fresh frozen tissue specimen in a sterile container; Frozen Stability: Ambient: Unacceptable Refrigerated: Stable up to 24 hours at 2–8 °C Frozen: Stable for 2 weeks at minus 20 °C Methodology: Polymerase Chain Reaction (PCR) Sequencing Days Performed: Once per week Reported: 1–2 weeks CPT: 87801 x 1	7/31/18
		Price: \$245.00 (non-discountable)	
CEBPA Mutation Analysis Marrow	CEBPAM	Note: This test was previously announced in the May 2018 Technical Update. Price: \$689.00 (non-discountable)	6/26/18
Cytomegalovirus Antiviral Drug Resistance by Sequencing	CYTSEQ	Special Information: Please submit most recent viral load and test date, if available. Heparinized specimens are unacceptable. This test is New York DOH approved. Clinical Limitation: This test may be unsuccessful if the plasma CMV DNA viral load is < 1,500 CMV DNA copies/mL of plasma. Clinical Information: Used to determine antiviral drug resistance to cidofovir, foscarnet sodium, and ganciclovir. Codons 457–630 of the UL97 gene and codons 393–1000 of the UL54 gene are sequenced. Mutations associated with resistance to ganciclovir, cidofovir, and foscarnet are reported. Mutations in viral subpopulations below 20% of total may not be detected. Specimen Requirement: 4 mL plasma from an EDTA (lavender) tube; Minimum: 1.5 mL; Draw 2 tubes to ensure adequate volume; Separate plasma from cells within 8 hours of collection; Transfer 4 mL plasma into a standard aliquot tube; Submit most recent viral load and test date, if available; Frozen *OR* 4 mL plasma from an EDTA (white) plasma preparation tube (PPT); Minimum: 1.5 mL; Draw 2 tubes to ensure adequate volume; Separate plasma from cells within 8 hours of collection; Transfer 4 mL plasma into a standard aliquot tube; Submit most recent viral load and test date, if available; Frozen Stability: Ambient: 8 hours Refrigerated: 72 hours Frozen: 1 month Methodology: Polymerase Chain Reaction/Sequencing Days Performed: Sunday—Saturday Reported: 3–4 days CPT: 87910 x 1 Price: \$639.00 (non-discountable)	8/7/18

Test Name	Order Code	Change	Effective Date
FLT3 Tyrosine Kinase Domain Analysis Blood	F3TKD	Specimen Requirement: 4 mL whole blood in an EDTA (lavender) tube; Minimum: 1 mL; Ambient Stability: Ambient: 48 hours Refrigerated: 7 days Frozen: Unacceptable Methodology: Next Gen Sequencing Days Performed: 2 days per week Reported: 7 days CPT: 81246 x 1, G0452 x 1 Price: \$704.00 (non-discountable)	6/26/18
FLT3 Tyrosine Kinase Domain Mutation, Bone Marrow	F3TKDM	Note: This test was previously announced in the May 2018 Technical Update. Please note that the order code FLT3TM has been changed to F3TKDM. Price: \$704.00 (non-discountable)	6/26/18
High Sensitivity Total Testosterone	HSTSTO	Clinical Information: Reference Intervals verified from Kushnir MM, Blamires T, Rockwood AL, Roberts WL, Yue B, Erdogan E, et al. Liquid chromatographytandem mass spectrometry assay for androstenedione, dehydroepiandrosterone, and testosterone with pediatric and adult reference intervals. Clin Chem 2010;56:1138-47 Specimen Requirement: 2 mL serum from a plain no additive (red) tube; Minimum: 1 mL; Separate serum from cells within 2 hours of collection; Keep refrigerated for transport; Refrigerated *OR* 2 mL plasma from a sodium heparin (green) tube; Minimum: 1 mL; Separate plasma from cells within 2 hours of collection; Keep refrigerated for transport; Refrigerated Stability: Ambient: 4 days Refrigerated: 14 days Refrigerated: 14 days Refrigerated: 14 days Frozen: 6 months Methodology: Liquid Chromatography-Tandem Mass Spectrometry (LC-MS/MS) Reference Range: Female Premature (26–28 Weeks): 5–16 ng/dL Premature (31–35 Weeks): 5–22 ng/dL Newborn: 20–64 ng/dL 1–5 Months: < 20 ng/dL 6–24 Months: < 9 ng/dL 2–3 Years: < 30 ng/dL 6–7 Years: < 7 ng/dL 8–9 Years: 1–11 ng/dL 10–11 Years: 3–32 ng/dL 12–13 Years: 6–50 ng/dL 14–15 Years: 6–50 ng/dL 18–59 Years: 9–55 ng/dL Premenopausal (18 Years and older): 9–55 ng/dL Premenopausal: 5–32 ng/dL Tanner Stage II: 2–17 ng/dL Tanner Stage II: 5–40 ng/dL Tanner Stage III: 10–63 ng/dL	7/31/18

Test Name	Order Code	Change	Effective Date
High Sensitivity Total Testosterone (continued from page 18)		Male Premature (26–28 Weeks): 59–125 ng/dL Premature (31–35 Weeks): 37–198 ng/dL Newborn: 75–400 ng/dL 1–5 Months: 14–363 ng/dL 6–24 Months: < 37 ng/dL 2–3 Years: < 15 ng/dL 4–5 Years: < 19 ng/dL 6–7 Years: < 13 ng/dL 8–9 Years: 2–8 ng/dL 10–11 Years: 2–165 ng/dL 12–13 Years: 3–619 ng/dL 14–15 Years: 31–733 ng/dL 16–17 Years: 158–826 ng/dL 18–39 Years: 300–1080 ng/dL 40–59 Years: 300–890 ng/dL 60 Years and older: 300–720 ng/dL Tanner Stage II: 2–15 ng/dL Tanner Stage III: 10–851 ng/dL Tanner Stage III: 10–851 ng/dL Tanner Stage IV-V: 162–847 ng/dL Days Performed: Monday–Friday Reported: 1–5 days CPT: 84403 x 1	
JAK2 Exon 12-15 Mutation Detection Bone Marrow	JAK2NM	Note: This test was previously announced in the May 2018 Technical Update. Price: \$561.00 (non-discountable)	6/26/18
JAK2 V617F Mutation Detection Bone Marrow	JAK2M	Note: This test was previously announced in the May 2018 Technical Update. Price: \$561.00 (non-discountable)	6/26/18
MPL Mutation Analysis Marrow	MPLM	Note: This test was previously announced in the May 2018 Technical Update. Price: \$942.00 (non-discountable)	6/26/18
Myeloproliferative Neoplasm Panel Marrow	MPNM	Note: This test was previously announced in the May 2018 Technical Update. Price: \$719.00 (non-discountable)	6/26/18
Myeloproliferative Neoplasm Panel Peripheral Blood	MPNP	Note: This test was previously announced in the May 2018 Technical Update. Price: \$719.00 (non-discountable)	6/26/18
NPM1 Mutation Detection Bone Marrow	NPM1M	Note: This test was previously announced in the May 2018 Technical Update. Price: \$691.00 (non-discountable)	6/26/18

Test Name	Order Code	Change	Effective Date
Toxicology Screen with Confirmation, Urine	UTOXRF	Includes: Ethanol, Urine Phencyclidine, Urine Benzodiazepines, Ur Cocaine, Urine Amphetamines, Urine Cannabinoids, Urine Opiates, Urine Barbiturates, Urine Oxycodone, Urine	7/31/18
		Special Information: Urine sent for confirmation for all components not reported as negative.	
		Specimen Requirement: 5 mL random urine in a clean container; Minimum: 3 mL; Refrigerated	
		Stability: Ambient: Unacceptable Refrigerated: 5 days Frozen: Unacceptable	
		Methodology: Enzymatic Kinetic Interaction of Microparticles in a Solution	
		Days Performed: Sunday–Saturday	
		Reported: 8 hours	
		CPT: 80307 x 1	
		Price: \$195.00	
Urogenital Ureaplasma and Mycoplasma Species by PCR	URMPCR	Special Information: Specimen source required. This test is New York DOH approved. Clinical Information: This test detects and speciates Ureaplasma parvum, Ureaplasma urealyticum, Mycoplasma hominis, and Mycoplasma genitalium; consider ordering for cases of non-gonococcal urethritis. A negative result does not rule out the presence of PCR inhibitors in the patient specimen or test-specific nucleic acid in concentrations below the level of detection by this test. Specimen Requirement: Genital swab in Viral Transport Media (VTM); Minimum: 0.5 mL; Transfer genital swab to VTM; Specimen source required; Frozen *OR* 1 mL random urine in Viral Transport Media (VTM); Minimum: 0.5 mL; Transfer 1 mL urine to VTM; Specimen source required; Frozen *OR* Cervical specimen collected using ThinPrep Pap Test Collection kit; Minimum: 0.5 mL; Vortex ThinPrep PreservCyt solution and transfer 1 mL into a sterile container; Specimen source required; Frozen *OR* Vaginal specimen collected using ThinPrep Pap Test Collection kit; Minimum: 0.5 mL; Vortex ThinPrep PreservCyt solution and transfer 1 mL into a sterile container; Specimen source required; Frozen *OR* Vaginal specimen collected using ThinPrep Pap Test Collection kit; Minimum: 0.5 mL; Vortex ThinPrep PreservCyt solution and transfer 1 mL into a sterile container; Specimen source required; Frozen Stability: Ambient: 24 hours Refrigerated: 10 days Frozen: 3 months	8/9/18
		Methodology: Qualitative Polymerase Chain Reaction	
		Days Performed: Monday, Thursday	
		Reported: 3–6 days	
		CPT: 87798 x 1	
		Price: \$206.00 (non-discountable)	

Test Name	Order Code	Change	Effective Date
Very Long-Chain and Branched-Chain Fatty Acids Profile	FATLON	Includes: Pristanic Acid Phytanic Acid Ratio Pristanic Acid to Phytanic Acid C22-0 Behenic Acid C22-0 Tetracosanoic Acid Ratio C24-0 to C22-0 Ratio C26-0 to C22-0 Ratio C26-0 to C22-0 Special Information: CRITICAL FROZEN. Clinical information is needed for appropriate interpretation. Age, gender, diet (e.g., TPN therapy), drug therapy, and family history are also required. Patient Prep: Fasting specimen is preferred for adults. For infants and children, draw specimen prior to feeding or 2-3 hours after a meal. Must submit Biochemical Genetics Patient History Form with the specimen. Separate specimens must be submitted when multiple tests are ordered. Room temperature specimens greater than 24 hours and refrigerated specimens greater than 48 hours are unacceptable. Specimens exposed to more than one freeze/thaw cycle are not acceptable. This test is New York DOH approved. Clinical Information: This is an initial test to screen for disorders of peroxisomal biogenesis and/or function, including X-linked adrenoleukodystrophy and Zellweger syndrome. Specimen Requirement: 0.5 mL plasma from a sodium or lithium heparin (green) tube; Minimum: 0.2 mL; Patient Prep: Fasting specimen is preferred for adults; For infants and children, draw specimen prior to feeding or 2-3 hours after a meal; Separate plasma from cells and transfer into a standard aliquot tube; Freeze immediately; Must submit Biochemical Genetics Patient History Form with the specimen; Separate specimens must be submitted when multiple tests are ordered; Critical Frozen *OR* 0.5 mL plasma from an EDTA (lavender) tube; Minimum: 0.2 mL; Patient Prep: Fasting specimen is preferred for adults; For infants and children, draw specimen prior to feeding or 2-3 hours after a meal; Separate plasma from cells and transfer into a standard aliquot tube; Freeze immediately; Must submit Biochemical Genetics Patient History Form with the specimen; Separate specimens must be	8/2/18

Very Long-Chain and Branched-Chain	Test Name	Order Code	Change	Effective Date
CPT: 82726 x 1	and Branched-Chain Fatty Acids Profile (continued from		0–11 Months: 28.94–93.50 μmol/L 1–2 Years: 28.94–93.50 μmol/L 3–6 Years: 28.94–93.50 μmol/L 7 Years and older: 28.94–93.50 μmol/L C24:0 Tetracosanoic Acid 0–11 Months: 24.25–77.75 μmol/L 1–2 Years: 24.25–77.75 μmol/L 3–6 Years: 24.25–77.75 μmol/L 7 Years and older: 24.25–77.75 μmol/L 7 Years and older: 24.25–77.75 μmol/L C26:0 Hexacosanoic Acid 0–11 Months: 0.17–0.73 μmol/L 1–2 Years: 0.17–0.73 μmol/L 3–6 Years: 0.17–0.73 μmol/L 7 Years and older: 0.17–0.73 μmol/L Ratio C24:0 to C22:0 0–11 Months: 0.64–1.02 1–2 Years: 0.64–1.02 3–6 Years: 0.64–1.02 7 Years and older: 0.64–1.02 Ratio C26:0 to C22:0 0–11 Months: 0.003–0.015 1–2 Years: 0.003–0.015 7 Years and older: 0.003–0.015 7 Years and older: 0.003–0.015 7 Years and older: 0.003–0.015	

Fee Reductions

Test Name	Order Code	List Fee	CPT Code	Effective Date
Calprotectin, Fecal	CALPRO	\$115.00	83993	7/31/18
Norovirus Group 1 and 2 Detection by PCR	NORPCR	\$176.00 (non-discountable)	87798	Effective immediately

Discontinued Tests

Test Name	Order Code	Test Information	Effective Date
BK Virus PCR Qual, Urine	UBKQAL	This test will no longer be available. Suggest ordering BK Virus Quantitation, Urine (UBKQT).	8/2/18
Chlamydia trachomatis DFA	CHLDFA	This test will no longer be available. Suggest ordering Chlamydia Amplification, Genital, Rectal and Oral Specimens (CT) or Chlamydia trachomatis, NA Amplification, Ocular Specimens (CTNAAO).	6/25/18
Cytomegalovirus Antiviral Drug Resistance	CYTOSQ	This test will no longer be available. Suggest ordering Cytomegalovirus Antiviral Drug Resistance by Sequencing (CYTSEQ).	8/7/18
Long Chain Fatty Acids	LONFAT	This test will no longer be available. Suggest ordering Very Long-Chain and Branched-Chain Fatty Acids Profile (FATLON).	8/2/18
Mycoplasma hominis PCR	MYPCR	This test will no longer be available. Suggest ordering Urogenital Ureaplasma and Mycoplasma Species by PCR (URMPCR).	8/9/18
Oil Red O LC	OILRED	This test will no longer be available.	7/31/18
Oncologic CytoScan HD SNP Array	HDSNP	This test will no longer be available.	8/2/18
Toxicology Screen w/ confirmation, Urine	UTOXWC	This test will no longer be available. Suggest ordering Toxicology Screen with Confirmation, Urine (UTOXRF).	7/31/18
Universal Bacterial, Fungal, and AFB PCR	FABPCR	This test will no longer be available. Suggest ordering Bacterial PCR, Direct Specimen (BCTPCR), Universal PCR, Fungal (FUNPCR) and Universal PCR, Acid Fast Bacilli (AFBPCR).	7/31/18
Whipple's Disease DNA by PCR	WHIPDN	This test will no longer be available. Suggest ordering Tropheryma whipplei PCR (WHIPWB).	7/31/18